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**POLYNUCLEOTIDES ENCODING MEMBERS OF THE
HUMAN B LYMPHOCYTE ACTIVATION ANTIGEN B7
FAMILY AND POLYPEPTIDES ENCODED THEREBY**

FIELD OF THE INVENTION

5 The invention relates generally to polynucleotides and polypeptides encoded thereby, as well as vectors, antibodies and recombinant methods for producing the polypeptides and polynucleotides.

BACKGROUND OF THE INVENTION

10 The immune system of vertebrates is characterized by its ability to discriminate "self" from "non-self" and to mount an appropriate, selective response to pathogens and other potentially harmful agents. Cell types involved in immune responses include lymphocytes known as B cells and T cells. Interactions between B cells and T cells are important for the propagation of full immune responses.

15 T cells must be activated in order to effect an immune response. T cell activation is thought to require two signals: an antigen-specific signal and a signal that is not antigen-specific. T cells can become activated by binding to B cells, particularly B cells which are themselves activated.

20 B cell-mediated activation of T cells is thought to be mediated, at least in part, by B7 proteins, which are expressed on the surface of B cells. B7 proteins are members of the immunoglobulin superfamily. They bind to activated T lymphocytes and provide regulatory signals for T lymphocyte cell growth and activation. (See, e.g., "Immunobiology - The Immune System in Health and Disease", 1997, Third edition, chapter 7, Janeway, C.A. et al., eds., Garland Publishing Inc., New York.) Cell surface molecules on T cells which bind to B7 molecules include CTLA4 and CD28.

SUMMARY OF THE INVENTION

The present invention is based, in part, upon the discovery of BLAA polynucleotide sequences encoding novel members of the human B Lymphocyte Activation Antigen B7 family (henceforth "BLAA").

5 In one aspect, the present invention involves an isolated nucleic acid molecule encoding a BLAA. This nucleic acid molecule may be a nucleotide that encodes a polypeptide having a sequence that is at least 95% identical to SEQ ID NO:2 or SEQ ID NO:6. Alternatively, it may be the complement of such a nucleic acid molecule.

10 In another aspect, this invention involves an oligonucleotide that is less than 100 nucleotides in length and contains at least 6 contiguous nucleotides of SEQ ID NO:1, 3, 5, 7, 8, 9, 10, 11, or 12. This oligonucleotide contains at least 6 contiguous nucleotides of a complement of SEQ ID NO:1, 3, 5, 7, 8, 9, 10, 11, or 12.

15 Another aspect of this invention relates to a vector containing an isolated nucleic acid molecule encoding a BLAA, as described above. In one embodiment, this vector is an expression vector. In another embodiment, this vector contains a regulatory element that is operably linked to the isolated nucleic acid molecule.

A further aspect of this invention involves an isolated polypeptide that is at least 80% identical to a polypeptide having an amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, or SEQ ID NO:6. In another embodiment of this aspect of the invention, the isolated polypeptide is at least 80% homologous to a fragment of a polypeptide having an amino acid sequence of
20 SEQ ID NO:2, SEQ ID NO:4, or SEQ ID NO:6. In this embodiment, the fragment contains at least 6 contiguous amino acids of SEQ ID NO:2, SEQ ID NO:4, or SEQ ID NO:6. Other embodiments of this aspect of the invention require that the isolated polypeptides of the invention be at least 80% homologous to a derivative, analog, or homolog of a polypeptide
25 having an amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, or SEQ ID NO:6. Yet another embodiment involves an isolated polypeptide that is at least 80% identical to a naturally occurring allelic variant of a polypeptide having an amino acid sequence of SEQ ID NO:2, SEQ ID NO:4, or SEQ ID NO:6. This embodiment requires that the polypeptide be encoded by a nucleic acid molecule capable of hybridizing to a nucleic acid molecule of SEQ
30 ID NO:1, SEQ ID NO:3, or SEQ ID NO:5, under stringent conditions.

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Another embodiment of this invention involves an antibody that selectively binds to the polypeptide(s) described above. Further embodiments of this invention provide methods for (a) producing such polypeptide(s) by culturing a host cell under conditions in which the nucleic acid molecule is expressed; (b) detecting the presence of the polypeptide(s) in a sample
5 by contacting the sample with a compound that selectively binds to the polypeptide(s) of the invention; and (c) modulating the activity of such polypeptide(s) by contacting a cell sample containing the polypeptide(s) of the instant invention with a compound that binds to the polypeptide(s) to modulate its activity.

Additionally, a method of detecting the presence of one of the nucleic acid molecules
10 of this invention is also provided. This method involves contacting a sample with a compound that selectively binds to the polypeptide(s) of this invention and determining whether the compound bound to the polypeptide(s) is present in the sample.

Other embodiments of this invention include methods of treating or preventing immune response-associated disorders and therapeutic or prophylactic pharmaceutical
15 compositions. Additionally, another aspect of this invention involves a kit containing therapeutically or prophylactically effective amounts of the pharmaceutical compositions provided for in the instant invention.

Another aspect of this invention involves the use of a therapeutic in the manufacture of a medicament for treating diseases associated with immune-response disorders. Another
20 embodiment of this invention involves a method for screening for a modulator of activity, latency or predisposition to an immune response-associated disorder. In this embodiment, a test compound is administered to an animal at an increased risk for an immune response-associated disorder, measuring the expression of the polypeptide of the invention in the test animal, measuring polypeptide expression in a control animal, and comparing the expression
25 in both animals.

Other embodiments of this invention include methods for determining the presence of or predisposition to a disease associate with altered levels of a BLAA polypeptide or a BLAA nucleic acid by measuring the amount of BLAA polypeptide or nucleic acid in a sample and comparing it to the amount present in a control sample. Yet other embodiments of this
30 invention involve methods of treating a pathological state in a mammal by administering a

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BLAA polypeptide or a BLAA nucleic acid in amounts sufficient to alleviate the pathological state.

Unless otherwise defined, all technical and scientific terms used herein have the same meaning as commonly understood by one of ordinary skill in the art to which this invention
5 pertains. Although methods and materials similar or equivalent to those described herein can be used in the practice or testing of the present invention, suitable methods and materials are described below. All publications, patent applications, patents, and other references
mentioned herein are incorporated by reference in their entirety. In case of conflict, the present specification, including definitions, will control. In addition, the materials, methods,
10 and examples are illustrative only and are not intended to be limiting.

Other features and advantages of the invention will be apparent from the following detailed description and claims.

BRIEF DESCRIPTION OF THE DRAWINGS

Figure 1 is a representation of a 2691 nucleotide human cDNA sequence (SEQ ID NO: 1), and
15 an amino acid sequence of the encoded polypeptide (SEQ ID NO:2).

Figure 2 is a representation of a 2885 nucleotide human cDNA sequence (SEQ ID NO: 3), and
an amino acid sequence of the encoded polypeptide (SEQ ID NO:4).

20 Figure 3 is a representation of a 2229 nucleotide human cDNA sequence (SEQ ID NO: 5), and
an amino acid sequence of the encoded polypeptide (SEQ ID NO:6).

25 Figure 4 shows the ClustalW Alignment of the BLAA polypeptide of SEQ ID NO: 6 as compared to several homologous B7 proteins and indicates the conserved regions by dark highlighting and conservative amino acid substitutions by lighter highlighting.

Figure 5 depicts the hydrophobicity plot for the polypeptide of SEQ ID NO: 6.

DETAILED DESCRIPTION OF THE INVENTION

The invention provides novel polynucleotides encoding polypeptides bearing sequence similarity to the B7 family of B lymphocyte antigens (BLAA). These cDNA sequences according to the invention, along with their encoded BLAA polypeptides are shown in Figures 1, 2 and 3.

The BLAA polypeptides are 20-25% identical and 40-45% similar to previously described members of the B7 family (*i.e.*, B7-1 and B7-2). As discussed in detail below, several motifs of the BLAA polypeptides are highly conserved with B7 polypeptides. Additionally, comparable similarity to human, mouse, and bovine butyrophilin as well as human myelin oligodendrocyte glycoprotein (MOG) has been observed.

The polypeptides of this invention have two (SEQ ID NO: 2), three (SEQ ID NO: 4), or four (SEQ ID NO: 6) Ig-like domains, followed by a transmembrane domain and a 44 amino acid cytoplasmic domain. This is consistent with previously described members of the B7 protein family, which are known to be localized on the cell surface (see, e.g., Selvakumar et al., Immunogenetics 36:175-181 (1992) and Linsley et al., Protein Sci 3:1341-1343 (1994)).

As used herein, the term "human B lymphocyte activation antigen B7-like activity" encompasses binding to activated T lymphocytes and providing regulatory signals for T lymphocyte cell growth and activation.

Accordingly, the polypeptides of the instant invention are novel members of the B7 protein family. The nucleic acid molecules and polypeptides of the present invention can be used to modulate immune responses. As such, they are useful in potential therapeutic applications implicated in the treatment of infectious diseases, cancers, autoimmune disorders and complications associated with graft vs. host disease in organ transplantation, and/or in similar immune system pathologies and disorders. For example, a cDNA encoding a polypeptide of the instant invention may be useful in gene therapy. Alternatively, a BLAA polypeptide can be administered directly to treat immune disorders. The nucleic acids, polypeptides and antibodies of the instant invention may be further useful in diagnostic and therapeutic applications.

The BLAA polypeptides contain numerous regions and/or single amino acids that are conserved at corresponding position in other homologous B7 proteins. For example, Figure 4 depicts the ClustalW alignment of the polypeptide of SEQ ID NO: 6. Conserved regions

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include, e.g., amino acids 16, 42, 46, 48, 50, 52, 59, 60, 63, 67-68, 75, 82, 89, 92-93, 107-109, 116, 118, 122, 134, 136, 138, 140, 143, 145, 152, 164-167, 170, 172-173, 187, 191, 194, 198, 210, 217, 223, 242 and 310. In addition, there are also regions with conservative amino acid substitutions as defined in the section "Conservative Mutations". By "conservative amino acid
 5 substitutions" is meant amino acids having similar side chains. Such substitutions are found, e.g., at amino acids 14, 25, 38, 65, 71, 79, 84, 106, 112, 114, 119, 123-124, 127-128, 142, 148-149, 161, 163, 171, 202, 212, 217, 221-223, 264, 266, 278, 283-284, 291, 305 and 335.

The BLAA polypeptides include hydrophilic and hydrophobic regions. As shown in Figure 5, the polypeptide of SEQ ID NO: 6 contains both hydrophilic and hydrophobic
 10 portions. Hydrophobic regions include from about amino acid 1 to about amino acid 75; from about amino acid 110 to about amino acid 150; from about amino acid 200 to about amino acid 225; from about amino acid 250 to about amino acid 290; from about amino acid 310 to about amino acid 380; from about amino acid 420 to about amino acid 440; and from about amino acid 460 to about amino acid 500. Conversely, the hydrophilic regions include from
 15 about amino acid 75 to about amino acid 110; from about amino acid 150 to about amino acid 200; from about amino acid 225 to about amino acid 250; from about amino acid 290 to about amino acid 310; from about amino acid 380 to about amino acid 420; from about amino acid 440 to about amino acid 534. Accordingly, these regions are useful for designing epitopes or selecting antigens.

20 For ease of reference, the novel polynucleotide and polypeptide sequences of the present invention shall be referred to collectively as BLAA (for B Lymphocyte Activation Antigen). A summary of the BLAA nucleic acid and polypeptide sequences of the present invention is provided in Table 1.

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Table 1: Sequences and Corresponding SEQ ID Numbers

Sequence Identifier Number	Description
SEQ ID NO: 1	2691 nucleotide human cDNA sequence
SEQ ID NO: 2	340 amino acid polypeptide encoded by nucleotides 111-1130 (SEQ ID NO: 7) of SEQ ID NO: 1
SEQ ID NO: 3	2885 nucleotide human cDNA sequence
SEQ ID NO: 4	441 amino acid polypeptide encoded by nucleotides 2-1324 (SEQ ID NO: 9) of SEQ ID NO: 3
SEQ ID NO: 5	2229 nucleotide human cDNA sequence
SEQ ID NO: 6	534 amino acid polypeptide encoded by nucleotides 60-1661 (SEQ ID NO: 11) of SEQ ID NO: 5
SEQ ID NO: 7	Open reading frame extending from nucleotide 110 to 1130 of SEQ ID NO: 1
SEQ ID NO: 8	3' non-translated region ("NTR") extending from nucleotide 1130 to 2691 of SEQ ID NO: 1
SEQ ID NO: 9	Open reading frame extending from nucleotide 2 to 1324 of SEQ ID NO: 3
SEQ ID NO: 10	3' NTR extending from nucleotide 1325 to 2885 of SEQ ID NO: 3
SEQ ID NO: 11	Open reading frame extending from nucleotide 60 to 1661 of SEQ ID NO: 5
SEQ ID NO: 12	3' NTR extending from nucleotide 1662 to 2229 of SEQ ID NO: 7

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As used herein, "identical" residues correspond to those residues in a comparison between two sequences where the equivalent nucleotide base or amino acid residue in an alignment of two sequences is the same residue. Residues are "similar" when the comparisons between two sequences in an alignment show that residues in an equivalent position in a comparison are either the same amino acid or a conservatively substituted amino acid as defined below in the section "Conservative Mutations".

Nucleic Acids

One aspect of the invention pertains to isolated BLAA nucleic acid molecules that encode BLAA polypeptides or biologically active portions thereof. BLAA nucleic acid molecules also include nucleic acid fragments sufficient for use as hybridization probes to identify BLAA-encoding nucleic acids (*e.g.*, BLAA mRNA) and fragments for use as PCR primers for the amplification or mutation of BLAA nucleic acid molecules. As used herein, the term "nucleic acid molecule" is intended to include DNA molecules (*e.g.*, cDNA or genomic DNA), RNA molecules (*e.g.*, mRNA), analogs of the DNA or RNA generated using nucleotide analogs, and derivatives, fragments and homologs thereof. The nucleic acid molecule can be single-stranded or double-stranded, but preferably is double-stranded DNA.

"Probes" refer to nucleic acid sequences of variable length, preferably between at least about 10 nucleotides (nt), 100 nt, or as many as about, *e.g.*, 6,000 nt, depending on use. Probes are used in the detection of identical, similar, or complementary nucleic acid sequences. Longer length probes are usually obtained from a natural or recombinant source and are highly specific and much slower to hybridize than oligomers. Probes may be single- or double-stranded and designed to have specificity in PCR, membrane-based hybridization technologies, or ELISA-like technologies.

An "isolated" nucleic acid molecule is one that is separated from other nucleic acid molecules which are present in the natural source of the nucleic acid. Preferably, an "isolated" nucleic acid is free of sequences which naturally flank the nucleic acid (*i.e.*, sequences located at the 5' and 3' ends of the nucleic acid) in the genomic DNA of the organism from which the nucleic acid is derived. For example, in various embodiments, the isolated BLAA nucleic acid molecule can contain less than about 5 kb, 4 kb, 3 kb, 2 kb, 1 kb, 0.5 kb or 0.1 kb of nucleotide sequences which naturally flank the nucleic acid molecule in genomic DNA of the cell from

which the nucleic acid is derived. Moreover, an "isolated" nucleic acid molecule, such as a cDNA molecule, can be substantially free of other cellular material or culture medium when produced by recombinant techniques, or of chemical precursors or other chemicals when chemically synthesized.

5 A nucleic acid molecule of the present invention, *e.g.*, a nucleic acid molecule having a nucleotide sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 3, SEQ ID NO: 5, SEQ ID NO: 7, SEQ ID NO: 8, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, and SEQ ID NO: 12, or a complement of any of these nucleotide sequences, can be isolated using standard molecular biology techniques and the sequence information provided
10 herein. Using all or a portion of the nucleic acid sequences of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12 as a hybridization probe, BLAA molecules can be isolated using standard hybridization and cloning techniques (*e.g.*, as described in Sambrook *et al.*, (eds.), MOLECULAR CLONING: A LABORATORY MANUAL 2nd Ed., Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY, 1989; and Ausubel, *et al.*, (eds.), CURRENT PROTOCOLS IN
15 MOLECULAR BIOLOGY, John Wiley & Sons, New York, NY, 1993.)

A nucleic acid of the invention can be amplified using cDNA, mRNA or, alternatively, genomic DNA, as a template and appropriate oligonucleotide primers according to standard PCR amplification techniques. The nucleic acid so amplified can be cloned into an appropriate vector and characterized by DNA sequence analysis. Furthermore,
20 oligonucleotides corresponding to BLAA nucleotide sequences can be prepared by standard synthetic techniques, *e.g.*, using an automated DNA synthesizer.

As used herein, the term "oligonucleotide" refers to a series of linked nucleotide residues, which oligonucleotide has a sufficient number of nucleotide bases to be used in a PCR reaction. A short oligonucleotide sequence may be based on, or designed from, a
25 genomic or cDNA sequence and is used to amplify, confirm, or reveal the presence of an identical, similar or complementary DNA or RNA in a particular cell or tissue. Oligonucleotides comprise portions of a nucleic acid sequence having about 10 nt, 50 nt, or 100 nt in length, preferably about 15 nt to 30 nt in length. In one embodiment, an oligonucleotide comprising a nucleic acid molecule less than 100 nt in length would further
30 comprise at least 6 contiguous nucleotides of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12 or a

complement thereof. Oligonucleotides may be chemically synthesized and may be used as probes.

In another embodiment, an isolated nucleic acid molecule of the invention comprises a nucleic acid molecule that is a complement of the nucleotide sequence shown in SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12. A nucleic acid molecule that is complementary to the nucleotide sequence shown in SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12 is one that is sufficiently complementary to the nucleotide sequence shown in SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12, that it can hydrogen bond with little or no mismatches to the nucleotide sequence shown in SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12, thereby forming a stable duplex.

As used herein, the term "complementary" refers to Watson-Crick or Hoogsteen base pairing between nucleotides units of a nucleic acid molecule. and the term "binding" means the physical or chemical interaction between two polypeptides or compounds or associated polypeptides or compounds or combinations thereof. Binding includes ionic, non-ionic, Van der Waals, hydrophobic interactions, etc. A physical interaction can be either direct or indirect. Indirect interactions may be through or due to the effects of another polypeptide or compound. Direct binding refers to interactions that do not take place through, or due to, the effect of another polypeptide or compound, but instead are without other substantial chemical intermediates.

Moreover, the nucleic acid molecule of the invention can comprise only a portion of the nucleic acid sequence of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12, *e.g.*, a fragment that can be used as a probe or primer or a fragment encoding a biologically active portion of BLAA.

Fragments provided herein are defined as sequences of at least 6 (contiguous) nucleic acids or at least 4 (contiguous) amino acids, a length sufficient to allow for specific hybridization in the case of nucleic acids or for specific recognition of an epitope in the case of amino acids. respectively, and are at most some portion less than a full length sequence. Fragments may be derived from any contiguous portion of a nucleic acid or amino acid sequence of choice. Derivatives are nucleic acid sequences or amino acid sequences formed from the native compounds either directly or by modification or partial substitution. Analogs are nucleic acid sequences or amino acid sequences that have a structure similar to, but not

identical to, the native compound but differs from it in respect to certain components or side chains. Analogs may be synthetic or from a different evolutionary origin and may have a similar or opposite metabolic activity compared to wild type. Homologs are nucleic acid sequences or amino acid sequences of a particular gene that are derived from different species.

5 Derivatives and analogs may be full length or other than full length, if the derivative or analog contains a modified nucleic acid or amino acid, as described below. Derivatives or analogs of the nucleic acids or proteins of the invention include, but are not limited to, molecules comprising regions that are substantially homologous to the nucleic acids or proteins of the invention, in various embodiments, by at least about 30%, 50%, 70%, 80%, or
10 95% identity (with a preferred identity of 80-95%) over a nucleic acid or amino acid sequence of identical size or when compared to an aligned sequence in which the alignment is done by a computer homology program known in the art, or whose encoding nucleic acid is capable of hybridizing to the complement of a sequence encoding the aforementioned proteins under stringent, moderately stringent, or low stringent conditions. See *e.g.* Ausubel, *et al.*, CURRENT
15 PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, New York, NY, 1993, and below.

A "homologous nucleic acid sequence" or "homologous amino acid sequence," or variations thereof, refer to sequences characterized by a homology at the nucleotide level or amino acid level as discussed above. Homologous nucleotide sequences encode those sequences coding for isoforms of a BLAA polypeptide. Isoforms can be expressed in different
20 tissues of the same organism as a result of, for example, alternative splicing of RNA. Alternatively, isoforms can be encoded by different genes. In the present invention, homologous nucleotide sequences include nucleotide sequences encoding for a BLAA polypeptide of species other than humans, including, but not limited to, mammals, and thus can include, *e.g.*, mouse, rat, rabbit, dog, cat, cow, horse, and other organisms. Homologous
25 nucleotide sequences also include, but are not limited to, naturally occurring allelic variations and mutations of the nucleotide sequences set forth herein. A homologous nucleotide sequence does not, however, include a nucleotide sequence encoding a human BLAA polypeptide. Homologous nucleic acid sequences include those nucleic acid sequences that encode conservative amino acid substitutions (see below) in SEQ ID NO: 2, SEQ ID NO: 4, or
30 SEQ ID NO: 6, as well as a polypeptide having BLAA activity. Biological activities of the

BLAA proteins are described below. A homologous amino acid sequence does not encode the amino acid sequence of a human BLAA polypeptide.

A BLAA polypeptide can be encoded by an open reading frame ("ORF") of a BLAA nucleic acid, as described herein. For example, the invention includes a nucleic acid sequence comprising the stretch of nucleic acid sequences of SEQ ID NO: 7, SEQ ID NO: 9, and SEQ ID NO: 11 that comprise the ORFs of the instant invention and encode a polypeptide of SEQ ID NO: 2, SEQ ID NO: 4, and SEQ ID NO: 6, respectively.

An "open reading frame" ("ORF") corresponds to a nucleotide sequence that can potentially be translated into a polypeptide. A stretch of nucleic acids comprising an ORF is uninterrupted by a stop codon. An ORF that represents the coding sequence for a full protein begins with an ATG "start" codon and terminates with one of the three "stop" codons, namely, TAA, TAG, or TGA. For the purposes of this invention, an ORF may be any part of a coding sequence, with or without a start codon, a stop codon, or both. For an ORF to be considered as a good candidate for coding for a bona fide cellular protein, a minimum size requirement is often set, for example, a stretch of DNA that would encode a protein of 50 amino acids or more.

In one embodiment, a 1020 base pair (bp) ORF (SEQ ID NO: 7) includes nucleotide 111 to nucleotide 1130 of SEQ ID NO: 1. This ORF can be translated into a 340 amino acid polypeptide according to SEQ ID NO: 2. In another embodiment, a 1323 bp ORF (SEQ ID NO: 9) includes nucleotide 2 to nucleotide 1324 of SEQ ID NO: 3. This ORF can be translated into a 441 amino acid polypeptide according to SEQ ID NO: 4. In another embodiment, a 1601 bp ORF (SEQ ID NO: 11) includes nucleotide 60 to nucleotide 1661 of SEQ ID NO: 5. This ORF could be translated into a 534 amino acid polypeptide according to SEQ ID NO: 6.

The nucleotide sequence determined from the cloning of the human BLAA gene allows for the generation of probes and primers designed for use in identifying and/or cloning BLAA homologues in other cell types, *e.g.* from other tissues, as well as BLAA homologues from other mammals. The probe/primer typically comprises substantially purified oligonucleotide. The oligonucleotide typically comprises a region of nucleotide sequence that hybridizes under stringent conditions to at least about 12, 25, 50, 100, 150, 200, 250, 300, 350 or 400

consecutive sense strand nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12; or an anti-sense strand nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12; or of a naturally occurring mutant of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12.

Probes based on a human BLAA nucleotide sequence can be used to detect transcripts
 5 or genomic sequences encoding the same or homologous proteins. In various embodiments, the probe further comprises a label group attached thereto, *e.g.* the label group can be a radioisotope, a fluorescent compound, an enzyme, or an enzyme co-factor. Such probes can be used as a part of a diagnostic test kit for identifying cells or tissue which misexpress a BLAA protein, such as by measuring a level of a BLAA-encoding nucleic acid in a sample of cells
 10 from a subject *e.g.*, detecting BLAA mRNA levels or determining whether a genomic BLAA gene has been mutated or deleted.

"A polypeptide having a biologically active portion of BLAA" refers to polypeptides exhibiting activity similar, but not necessarily identical to, an activity of a polypeptide of the present invention, including mature forms, as measured in a particular biological assay, with or
 15 without dose dependency. A nucleic acid fragment encoding a "biologically active portion of BLAA" can be prepared by isolating a portion of SEQ ID NO: 1, 3, 5, 7, 9, or 11, that encodes a polypeptide having a BLAA biological activity (the biological activities of the BLAA proteins are described below), expressing the encoded portion of BLAA polypeptide (*e.g.*, by recombinant expression *in vitro*) and assessing the activity of the encoded portion of BLAA.

20 BLAA variants

The invention further encompasses nucleic acid molecules that differ from the nucleotide sequence shown in SEQ ID NO: 1, 3, 5, 7, 9, or 11, due to degeneracy of the genetic code and thus encode the same BLAA protein as that encoded by the nucleotide sequence shown in SEQ ID NO: 1, 3, 5, 7, 9, or 11. In another embodiment, an isolated
 25 nucleic acid molecule of the invention has a nucleotide sequence encoding a protein having an amino acid sequence shown in SEQ ID NO: 2, SEQ ID NO: 4 or SEQ ID NO: 6.

In addition to the human BLAA nucleotide sequence shown in SEQ ID NO: 1, 3, 5, 7, 9, or 11, it will be appreciated by those skilled in the art that DNA sequence polymorphisms that lead to changes in the amino acid sequences of BLAA may exist within a population (*e.g.*,
 30 the human population). Such genetic polymorphism in the BLAA gene may exist among

individuals within a population due to natural allelic variation. As used herein, the terms "gene" and "recombinant gene" refer to nucleic acid molecules comprising an open reading frame encoding a BLAA polypeptide, preferably a mammalian BLAA polypeptide. Such natural allelic variations can typically result in 1-5% variance in the nucleotide sequence of the BLAA gene. Any and all such nucleotide variations and resulting amino acid polymorphisms in BLAA that are the result of natural allelic variation and that do not alter the functional activity of BLAA are intended to be within the scope of the invention.

Moreover, nucleic acid molecules encoding BLAA polypeptides from other species, and thus that have a nucleotide sequence that differs from the human sequence of SEQ ID NO: 1, 3, 5, 7, 9, or 11, are intended to be within the scope of the invention. Nucleic acid molecules corresponding to natural allelic variants and homologues of the BLAA cDNAs of the invention can be isolated based on their homology to the human BLAA nucleic acids disclosed herein using the human cDNAs, or a portion thereof, as a hybridization probe according to standard hybridization techniques under stringent hybridization conditions. For example, a soluble human BLAA cDNA can be isolated based on its homology to human membrane-bound BLAA. Likewise, a membrane-bound human BLAA cDNA can be isolated based on its homology to soluble human BLAA.

Accordingly, in another embodiment, an isolated nucleic acid molecule of the invention is at least 6 nucleotides in length and hybridizes under stringent conditions to the nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12. In another embodiment, the nucleic acid is at least 10, 25, 50, 100, 250, 500, 1000, 1500, 2000, or more nucleotides in length. In another embodiment, an isolated nucleic acid molecule of the invention hybridizes to the coding region, for example SEQ ID NO: 1, 3, 5, 7, 9, or 11. As used herein, the term "hybridizes under stringent conditions" is intended to describe conditions for hybridization and washing under which nucleotide sequences at least 60% homologous to each other typically remain hybridized to each other.

Homologs (*i.e.*, nucleic acids encoding BLAA polypeptides derived from species other than human) or other related sequences (*e.g.*, paralogs) can be obtained by low, moderate or high stringency hybridization with all or a portion of the particular human sequence as a probe using methods well known in the art for nucleic acid hybridization and cloning.

As used herein, the phrase "stringent hybridization conditions" refers to conditions under which a probe, primer or oligonucleotide will hybridize to its target sequence, but to no other sequences. Stringent conditions are sequence-dependent and will be different in different circumstances. Longer sequences hybridize specifically at higher temperatures than shorter sequences. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength, pH and nucleic acid concentration) at which 50% of the probes complementary to the target sequence hybridize to the target sequence at equilibrium. Since the target sequences are generally present at excess, at T_m, 50% of the probes are occupied at equilibrium. Typically, stringent conditions will be those in which the salt concentration is less than about 1.0 M sodium ion, typically about 0.01 to 1.0 M sodium ion (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes, primers or oligonucleotides (*e.g.*, 10 nt to 50 nt) and at least about 60°C for longer probes, primers and oligonucleotides. Stringent conditions may also be achieved with the addition of destabilizing agents, such as formamide.

Stringent conditions are known to those skilled in the art and can be found in Ausubel *et al.*, (eds.), CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. Preferably, the conditions are such that sequences at least about 65%, 70%, 75%, 85%, 90%, 95%, 98%, or 99% homologous to each other typically remain hybridized to each other. A non-limiting example of stringent hybridization conditions are hybridization in a high salt buffer comprising 6X SSC, 50 mM Tris-HCl (pH 7.5), 1 mM EDTA, 0.02% PVP, 0.02% Ficoll, 0.02% BSA, and 500 mg/ml denatured salmon sperm DNA at 65°C, followed by one or more washes in 0.2X SSC, 0.01% BSA at 50°C. An isolated nucleic acid molecule of the invention that hybridizes under stringent conditions to the sequence of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12, corresponds to a naturally-occurring nucleic acid molecule. As used herein, a "naturally-occurring" nucleic acid molecule refers to an RNA or DNA molecule having a nucleotide sequence that occurs in nature (*e.g.*, encodes a natural protein).

In a second embodiment, a nucleic acid sequence that is hybridizable to the nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or

12, or fragments, analogs or derivatives thereof, under conditions of moderate stringency is provided. A non-limiting example of moderate stringency hybridization conditions are hybridization in 6X SSC, 5X Denhardt's solution, 0.5% SDS and 100 mg/ml denatured salmon sperm DNA at 55°C, followed by one or more washes in 1X SSC, 0.1% SDS at 37°C. Other
5 conditions of moderate stringency that may be used are well-known in the art. See, *e.g.*, Ausubel *et al.* (eds.), 1993, CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, NY, and Kriegler, 1990, GENE TRANSFER AND EXPRESSION, A LABORATORY MANUAL, Stockton Press, NY.

In a third embodiment, a nucleic acid that is hybridizable to the nucleic acid molecule
10 comprising the nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12, or fragments, analogs or derivatives thereof, under conditions of low stringency, is provided. A non-limiting example of low stringency hybridization conditions are hybridization in 35% formamide, 5X SSC, 50 mM Tris-HCl (pH 7.5), 5 mM EDTA, 0.02% PVP, 0.02% Ficoll, 0.2% BSA, 100 mg/ml denatured salmon sperm DNA, 10% (wt/vol) dextran sulfate at 40°C, followed by one
15 or more washes in 2X SSC, 25 mM Tris-HCl (pH 7.4), 5 mM EDTA, and 0.1% SDS at 50°C. Other conditions of low stringency that may be used are well known in the art (*e.g.*, as employed for cross-species hybridizations). See, *e.g.*, Ausubel *et al.* (eds.), 1993, CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, NY, and Kriegler, 1990, GENE TRANSFER AND EXPRESSION, A LABORATORY MANUAL, Stockton Press, NY; Shilo and
20 Weinberg, 1981, *Proc Natl Acad Sci USA* 78: 6789-6792.

Conservative mutations

In addition to naturally-occurring allelic variants of the BLAA sequence that may exist in the population, the skilled artisan will further appreciate that changes can be introduced by mutation into the nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 9, or 11, thereby leading to
25 changes in the amino acid sequence of the encoded BLAA polypeptide, without altering the functional ability of the BLAA polypeptide. For example, nucleotide substitutions leading to amino acid substitutions at "non-essential" amino acid residues can be made in the sequence of SEQ ID NO: 1, 3, 5, 7, 9, or 11. A "non-essential" amino acid residue is a residue that can be altered from the wild-type sequence of BLAA without altering the biological activity, whereas
30 an "essential" amino acid residue is required for biological activity. For example, amino acid

residues that are conserved among the BLAA polypeptides of the present invention, are predicted to be particularly unamenable to alteration. (See Figure 4 and discussion above).

In addition, amino acid residues that are conserved among family members of the BLAA polypeptides of the present invention, as indicated by the alignment presented in Figure 4, and as described above, are also predicted to be particularly unamenable to alteration. Other amino acid residues, however, (e.g., those that are not conserved or only semi-conserved among members of the BLAA polypeptides) may not be essential for activity and thus are likely to be amenable to alteration.

Another aspect of the invention pertains to nucleic acid molecules encoding BLAA polypeptides that contain changes in amino acid residues that are not essential for activity. Such BLAA polypeptides differ in amino acid sequence from SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6, yet retain biological activity. In one embodiment, the isolated nucleic acid molecule comprises a nucleotide sequence encoding a polypeptide, wherein the polypeptide comprises an amino acid sequence at least about 45% homologous to the amino acid sequence of SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6. Preferably, the polypeptide encoded by the nucleic acid molecule is at least about 60% homologous to SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6, more preferably at least about 70% homologous to SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6, still more preferably at least about 80% homologous to SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6, even more preferably at least about 90% homologous to SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6, and most preferably at least about 95% homologous to SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6.

An isolated nucleic acid molecule encoding a BLAA polypeptide homologous to the polypeptide of SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6 can be created by introducing one or more nucleotide substitutions, additions or deletions into the nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 9, or 11 such that one or more amino acid substitutions, additions or deletions are introduced into the encoded polypeptide.

Mutations can be introduced into SEQ ID NO: 1, 3, 5, 7, 9, or 11 by standard techniques, such as site-directed mutagenesis and PCR-mediated mutagenesis. Preferably, conservative amino acid substitutions are made at one or more predicted non-essential amino acid residues. A "conservative amino acid substitution" is one in which the amino acid residue

is replaced with an amino acid residue having a similar side chain. Families of amino acid residues having similar side chains have been defined in the art. These families include amino acids with basic side chains (*e.g.*, lysine, arginine, histidine), acidic side chains (*e.g.*, aspartic acid, glutamic acid), uncharged polar side chains (*e.g.*, glycine, asparagine, glutamine, serine, threonine, tyrosine, cysteine), nonpolar side chains (*e.g.*, alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan), beta-branched side chains (*e.g.*, threonine, valine, isoleucine) and aromatic side chains (*e.g.*, tyrosine, phenylalanine, tryptophan, histidine). Thus, a predicted nonessential amino acid residue in BLAA is replaced with another amino acid residue from the same side chain family. Alternatively, in another embodiment, mutations can be introduced randomly along all or part of a BLAA coding sequence, such as by saturation mutagenesis, and the resultant mutants can be screened for BLAA biological activity to identify mutants that retain activity. Following mutagenesis of SEQ ID NO: 1, 3, 5, 7, 9, or 11, the encoded polypeptide can be expressed by any recombinant technology known in the art and the activity of the polypeptide can be determined.

In one embodiment, a mutant BLAA polypeptide can be assayed for (1) the ability to form protein:protein interactions with other BLAA polypeptides, other cell-surface proteins, or biologically active portions thereof, (2) complex formation between a mutant BLAA polypeptide and a BLAA ligand, *e.g.*, CTLA4; (3) the ability of a mutant BLAA polypeptide to bind to an intracellular target protein or biologically active portion thereof (*e.g.* avidin proteins).

In yet another embodiment, a mutant BLAA can be assayed for the ability to perform immunoglobulin superfamily member activities, such as, (i) complex formation between a BLAA polypeptide and activated T lymphocytes; (ii) interaction of a BLAA polypeptide with a protein having substantial homology to the human B lymphocyte activation antigen B7 family of proteins; (iii) interaction of a BLAA polypeptide with a human B lymphocyte activation antigen B7 family member protein; and (iv) interaction of a BLAA polypeptide with other proteins. In yet another embodiment, a BLAA activity is at least one or more of the following activities: (i) modulation of immunoglobulin superfamily-related protein activity; (ii) regulation of T lymphocyte growth; and (iii) regulation of T lymphocyte activation.

Antisense

Another aspect of the invention pertains to isolated antisense nucleic acid molecules that are hybridizable to or complementary to the nucleic acid molecule comprising the nucleotide sequence of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12, or fragments, analogs or derivatives thereof. An "antisense" nucleic acid comprises a nucleotide sequence that is complementary to a "sense" nucleic acid encoding a protein, *e.g.*, complementary to the coding strand of a double-stranded cDNA molecule or complementary to an mRNA sequence. In specific aspects, antisense nucleic acid molecules are provided that comprise a sequence complementary to at least about 10, 25, 50, 100, 250 or 500 nucleotides or an entire BLAA coding strand, or to only a portion thereof. Nucleic acid molecules encoding fragments, homologs, derivatives and analogs of a BLAA polypeptide of SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6 or antisense nucleic acids complementary to a BLAA nucleic acid sequence of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12, are additionally provided.

In one embodiment, an antisense nucleic acid molecule is antisense to a "coding region" of the coding strand of a nucleotide sequence encoding BLAA. The term "coding region" refers to the region of the nucleotide sequence comprising codons which are translated into amino acid residues (*e.g.*, the coding region of human BLAA corresponds to nucleotides 111-1130 of SEQ ID NO: 1 (SEQ ID NO: 7), nucleotides 2-1324 of SEQ ID NO: 3 (SEQ ID NO: 9), or nucleotides 60-1601 of SEQ ID NO: 5 (SEQ ID NO: 11)). In another embodiment, the antisense nucleic acid molecule is antisense to a "noncoding region" of the coding strand of a nucleotide sequence encoding BLAA. The term "noncoding region" refers to 5' and 3' sequences which flank the coding region that are not translated into amino acids (*i.e.*, also referred to as 5' and 3' untranslated regions). Examples of "noncoding regions" include the non-translated regions ("NTRs") of SEQ ID NO: 8, SEQ ID NO: 10, and SEQ ID NO: 12.

Given the coding strand sequences encoding BLAA disclosed herein (*e.g.*, SEQ ID NO: 5, SEQ ID NO: 7, and SEQ ID NO: 9), antisense nucleic acids of the invention can be designed according to the rules of Watson and Crick or Hoogsteen base pairing. The antisense nucleic acid molecule can be complementary to the entire coding region of BLAA mRNA, but more preferably is an oligonucleotide that is antisense to only a portion of the coding or noncoding region of BLAA mRNA. For example, the antisense oligonucleotide can be

complementary to the region surrounding the translation start site of BLAA mRNA. An antisense oligonucleotide can be, for example, about 5, 10, 15, 20, 25, 30, 35, 40, 45 or 50 nucleotides in length. An antisense nucleic acid of the invention can be constructed using chemical synthesis or enzymatic ligation reactions using procedures known in the art. For example, an antisense nucleic acid (*e.g.*, an antisense oligonucleotide) can be chemically synthesized using naturally occurring nucleotides or variously modified nucleotides designed to increase the biological stability of the molecules or to increase the physical stability of the duplex formed between the antisense and sense nucleic acids, *e.g.*, phosphorothioate derivatives and acridine substituted nucleotides can be used.

Examples of modified nucleotides that can be used to generate the antisense nucleic acid include: 5-fluorouracil, 5-bromouracil, 5-chlorouracil, 5-iodouracil, hypoxanthine, xanthine, 4-acetylcytosine, 5-(carboxyhydroxymethyl) uracil, 5-carboxymethylaminomethyl-2-thiouridine, 5-carboxymethylaminomethyluracil, dihydrouracil, beta-D-galactosylqueosine, inosine, N6-isopentenyladenine, 1-methylguanine, 1-methylinosine, 2,2-dimethylguanine, 2-methyladenine, 2-methylguanine, 3-methylcytosine, 5-methylcytosine, N6-adenine, 7-methylguanine, 5-methylaminomethyluracil, 5-methoxyaminomethyl-2-thiouracil, beta-D-mannosylqueosine, 5'-methoxycarboxymethyluracil, 5-methoxyuracil, 2-methylthio-N6-isopentenyladenine, uracil-5-oxyacetic acid (*v*), wybutoxosine, pseudouracil, queosine, 2-thiocytosine, 5-methyl-2-thiouracil, 2-thiouracil, 4-thiouracil, 5-methyluracil, uracil-5-oxyacetic acid methylester, uracil-5-oxyacetic acid (*v*), 5-methyl-2-thiouracil, 3-(3-amino-3-N-2-carboxypropyl) uracil, (acp3)_w, and 2,6-diaminopurine. Alternatively, the antisense nucleic acid can be produced biologically using an expression vector into which a nucleic acid has been subcloned in an antisense orientation (*i.e.*, RNA transcribed from the inserted nucleic acid will be of an antisense orientation to a target nucleic acid of interest, described further in the following subsection).

The antisense nucleic acid molecules of the invention are typically administered to a subject or generated *in situ* such that they hybridize with or bind to cellular mRNA and/or genomic DNA encoding a BLAA polypeptide to thereby inhibit expression of the protein, *e.g.*, by inhibiting transcription and/or translation. The hybridization can be by conventional nucleotide complementarity to form a stable duplex, or, for example, in the case of an

antisense nucleic acid molecule that binds to DNA duplexes, through specific interactions in the major groove of the double helix. An example of a route of administration of antisense nucleic acid molecules of the invention includes direct injection at a tissue site. Alternatively, antisense nucleic acid molecules can be modified to target selected cells and then administered systemically. For example, for systemic administration, antisense molecules can be modified such that they specifically bind to receptors or antigens expressed on a selected cell surface, e.g., by linking the antisense nucleic acid molecules to peptides or antibodies that bind to cell surface receptors or antigens. The antisense nucleic acid molecules can also be delivered to cells using the vectors described herein. To achieve sufficient intracellular concentrations of antisense molecules, vector constructs in which the antisense nucleic acid molecule is placed under the control of a strong pol II or pol III promoter are preferred.

In yet another embodiment, the antisense nucleic acid molecule of the invention is an α -anomeric nucleic acid molecule. An α -anomeric nucleic acid molecule forms specific double-stranded hybrids with complementary RNA in which, contrary to the usual β -units, the strands run parallel to each other (Gaultier *et al.* (1987) *Nucleic Acids Res* 15: 6625-6641). The antisense nucleic acid molecule can also comprise a 2'-o-methylribonucleotide (Inoue *et al.* (1987) *Nucleic Acids Res* 15: 6131-6148) or a chimeric RNA-DNA analogue (Inoue *et al.* (1987) *FEBS Lett* 215: 327-330).

Ribozymes and PNA moieties

Nucleic acid modifications include, by way of nonlimiting example, modified bases, and nucleic acids whose sugar phosphate backbones are modified or derivatized. These modifications are carried out at least in part to enhance the chemical stability of the modified nucleic acid, such that they may be used, for example, as antisense binding nucleic acids in therapeutic applications in a subject.

In one embodiment, an antisense nucleic acid of the invention is a ribozyme. Ribozymes are catalytic RNA molecules with ribonuclease activity that are capable of cleaving a single-stranded nucleic acid, such as an mRNA, to which they have a complementary region. Thus, ribozymes (e.g., hammerhead ribozymes (described in Haselhoff and Gerlach (1988) *Nature* 334:585-591)) can be used to catalytically cleave BLAA mRNA transcripts to thereby inhibit translation of BLAA mRNA. A ribozyme having specificity for a BLAA-encoding

nucleic acid can be designed based upon the nucleotide sequence of a BLAA cDNA disclosed herein (*i.e.*, SEQ ID NO: 1, 3, 5, 7, 9, or 11). For example, a derivative of a Tetrahymena L-19 IVS RNA can be constructed in which the nucleotide sequence of the active site is complementary to the nucleotide sequence to be cleaved in a BLAA-encoding mRNA. See, 5 *e.g.*, Cech *et al.* U.S. Pat. No. 4,987,071; and Cech *et al.* U.S. Pat. No. 5,116,742. Alternatively, BLAA mRNA can be used to select a catalytic RNA having a specific ribonuclease activity from a pool of RNA molecules. See, *e.g.*, Bartel *et al.*, (1993) *Science* 261:1411-1418.

Alternatively, BLAA gene expression can be inhibited by targeting nucleotide 10 sequences complementary to the regulatory region of the BLAA (*e.g.*, the BLAA promoter and/or enhancers) to form triple helical structures that prevent transcription of the BLAA gene in target cells. See generally, Helene. (1991) *Anticancer Drug Des.* 6: 569-84; Helene. *et al.* (1992) *Ann. N.Y. Acad. Sci.* 660:27-36; and Maher (1992) *Bioassays* 14: 807-15.

In various embodiments, the nucleic acids of BLAA can be modified at the base 15 moiety, sugar moiety or phosphate backbone to improve, *e.g.*, the stability, hybridization, or solubility of the molecule. For example, the deoxyribose phosphate backbone of the nucleic acids can be modified to generate peptide nucleic acids (see Hyrup *et al.* (1996) *Bioorg Med Chem* 4: 5-23). As used herein, the terms "peptide nucleic acids" or "PNAs" refer to nucleic acid mimics, *e.g.*, DNA mimics, in which the deoxyribose phosphate backbone is replaced by 20 a pseudopeptide backbone and only the four natural nucleobases are retained. The neutral backbone of PNAs has been shown to allow for specific hybridization to DNA and RNA under conditions of low ionic strength. The synthesis of PNA oligomers can be performed using standard solid phase peptide synthesis protocols as described in Hyrup *et al.* (1996) above; Perry-O'Keefe *et al.* (1996) *PNAS* 93: 14670-675.

25 PNAs of BLAA can be used in therapeutic and diagnostic applications. For example, PNAs can be used as antisense or antigene agents for sequence-specific modulation of gene expression by, *e.g.*, inducing transcription or translation arrest or inhibiting replication. PNAs of BLAA can also be used, *e.g.*, in the analysis of single base pair mutations in a gene by, *e.g.*, PNA directed PCR clamping; as artificial restriction enzymes when used in combination with

other enzymes, *e.g.*, S1 nucleases (Hyrup B. (1996) above); or as probes or primers for DNA sequence and hybridization (Hyrup *et al.* (1996), above; Perry-O'Keefe (1996), above).

In another embodiment, PNAs of BLAA can be modified, *e.g.*, to enhance their stability or cellular uptake, by attaching lipophilic or other helper groups to PNA, by the formation of PNA-DNA chimeras, or by the use of liposomes or other techniques of drug delivery known in the art. For example, PNA-DNA chimeras of BLAA can be generated that may combine the advantageous properties of PNA and DNA. Such chimeras allow DNA recognition enzymes, *e.g.*, RNase H and DNA polymerases, to interact with the DNA portion while the PNA portion would provide high binding affinity and specificity. PNA-DNA chimeras can be linked using linkers of appropriate lengths selected in terms of base stacking, number of bonds between the nucleobases, and orientation (Hyrup (1996) above). The synthesis of PNA-DNA chimeras can be performed as described in Hyrup (1996) above and Finn *et al.* (1996) *Nucl Acids Res* 24: 3357-63. For example, a DNA chain can be synthesized on a solid support using standard phosphoramidite coupling chemistry, and modified nucleoside analogs, *e.g.*, 5'-(4-methoxytrityl)amino-5'-deoxy-thymidine phosphoramidite, can be used between the PNA and the 5' end of DNA (Mag *et al.* (1989) *Nucl Acid Res* 17: 5973-88). PNA monomers are then coupled in a stepwise manner to produce a chimeric molecule with a 5' PNA segment and a 3' DNA segment (Finn *et al.* (1996) above). Alternatively, chimeric molecules can be synthesized with a 5' DNA segment and a 3' PNA segment. See. Petersen *et al.* (1975) *Bioorg Med Chem Lett* 5: 1119-11124.

In other embodiments, the oligonucleotide may include other appended groups such as peptides (*e.g.*, for targeting host cell receptors *in vivo*), or agents facilitating transport across the cell membrane (see, *e.g.*, Letsinger *et al.*, 1989, *Proc. Natl. Acad. Sci. U.S.A.* 86:6553-6556; Lemaitre *et al.*, 1987, *Proc. Natl. Acad. Sci.* 84:648-652; PCT Publication No. W088/09810) or the blood-brain barrier (see, *e.g.*, PCT Publication No. W089/10134). In addition, oligonucleotides can be modified with hybridization triggered cleavage agents (See, *e.g.*, Krol *et al.*, 1988, *BioTechniques* 6:958-976) or intercalating agents. (See, *e.g.*, Zon, 1988, *Pharm. Res.* 5: 539-549). To this end, the oligonucleotide may be conjugated to another molecule, *e.g.*, a peptide, a hybridization triggered cross-linking agent, a transport agent, a hybridization-triggered cleavage agent. etc.

BLAA polypeptides

As used herein, the terms "protein" and "polypeptide" are intended to be interchangeable. The novel polypeptides of the invention include the BLAA polypeptides whose sequence is provided in SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6. The invention also includes mutant or variant polypeptides any of whose residues may be changed from the corresponding residue shown in SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6 while still encoding a polypeptide that maintains its BLAA activities and physiological functions, or a functional fragment thereof. In the mutant or variant protein, up to 20% or more of the residues may be so changed.

In general, an BLAA variant that preserves BLAA-like function includes any variant in which residues at a particular position in the sequence have been substituted by other amino acids, and further include the possibility of inserting an additional residue or residues between two residues of the parent protein as well as the possibility of deleting one or more residues from the parent sequence. Any amino acid substitution, insertion, or deletion is encompassed by the invention. In favorable circumstances, the substitution is a conservative substitution as defined above.

One aspect of the invention pertains to isolated BLAA polypeptides, and biologically active portions thereof, or derivatives, fragments, analogs or homologs thereof. Also provided are polypeptide fragments suitable for use as immunogens to raise anti-BLAA antibodies. In one embodiment, native BLAA polypeptides can be isolated from cells or tissue sources by an appropriate purification scheme using standard protein purification techniques. In another embodiment, BLAA polypeptides are produced by recombinant DNA techniques. Alternative to recombinant expression, a BLAA protein or polypeptide can be synthesized chemically using standard peptide synthesis techniques.

An "isolated" or "purified" polypeptide or biologically active portion thereof is substantially free of cellular material or other contaminating proteins or polypeptides from the cell or tissue source from which the BLAA polypeptide is derived, or substantially free from chemical precursors or other chemicals when chemically synthesized. The language "substantially free of cellular material" includes preparations of BLAA polypeptide in which the polypeptide is separated from cellular components of the cells from which it is isolated or

recombinantly produced. In one embodiment, the language "substantially free of cellular material" includes preparations of BLAA polypeptide having less than about 30% (by dry weight) of non-BLAA protein (also referred to herein as a "contaminating protein"), more preferably less than about 20% of non-BLAA protein, still more preferably less than about 10% of non-BLAA protein, and most preferably less than about 5% non-BLAA protein.

When the BLAA polypeptide or biologically active portion thereof is recombinantly produced, it is also preferably substantially free of culture medium, *i.e.*, culture medium represents less than about 20%, more preferably less than about 10%, and most preferably less than about 5% of the volume of the protein preparation.

10 The language "substantially free of chemical precursors or other chemicals" includes preparations of BLAA polypeptide in which the polypeptide is separated from chemical precursors or other chemicals that are involved in the synthesis of the polypeptide. In one embodiment, the language "substantially free of chemical precursors or other chemicals" includes preparations of BLAA polypeptide having less than about 30% (by dry weight) of chemical precursors or non-BLAA chemical, more preferably less than about 20% chemical precursors or non-BLAA chemicals, still more preferably less than about 10% chemical precursors or non-BLAA chemicals, and most preferably less than about 5% chemical precursors or non-BLAA chemicals.

Biologically active portions of a BLAA polypeptide include peptides comprising amino acid sequences sufficiently homologous to or derived from the amino acid sequence of the BLAA polypeptides, *e.g.*, the amino acid sequence shown in SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6, that include fewer amino acids than the full length BLAA polypeptides, and exhibit at least one activity of a BLAA polypeptide. Typically, biologically active portions comprise a domain or motif with at least one activity of the BLAA polypeptide. A biologically active portion of a BLAA polypeptide can be a polypeptide which is, for example, 10, 25, 50, 100 or more amino acids in length.

The polypeptide disclosed in SEQ ID NO: 2 has two Ig-like domains, followed by a transmembrane domain and a 44 amino acid cytoplasmic domain. The polypeptide disclosed in SEQ ID NO: 4 has three Ig-like domains, followed by a transmembrane domain and a 44 amino acid cytoplasmic domain. The polypeptide disclosed in SEQ ID NO: 6 has four Ig-like

domains, followed by a transmembrane domain and a 44 amino acid cytoplasmic domain. Its hydrophobicity plot analysis is shown in Figure 5.

In an embodiment, the BLAA polypeptide has an amino acid sequence shown in SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6. In other embodiments, the BLAA polypeptide is substantially homologous to SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6 and retains the functional activity of the polypeptide of SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6, yet differs in amino acid sequence due to natural allelic variation or mutagenesis, as described in detail below. Accordingly, in another embodiment, the BLAA polypeptide is a polypeptide that comprises an amino acid sequence at least about 45% homologous to the amino acid sequence of SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6 and retains the functional activity of the BLAA polypeptides of SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6.

Determining homology between two or more sequences

To determine the percent homology of two amino acid sequences or of two nucleic acids, the sequences are aligned for optimal comparison purposes (*e.g.*, gaps can be introduced in the sequence of a first amino acid or nucleic acid sequence for optimal alignment with a second amino or nucleic acid sequence). The amino acid residues or nucleotides at corresponding amino acid positions or nucleotide positions are then compared. When a position in the first sequence is occupied by the same amino acid residue or nucleotide as the corresponding position in the second sequence, then the molecules are homologous at that position (*i.e.*, as used herein amino acid or nucleic acid "homology" is equivalent to amino acid or nucleic acid "identity").

The nucleic acid sequence homology may be determined as the degree of identity between two sequences. The homology may be determined using computer programs known in the art, such as GAP software provided in the GCG program package. See, Needleman and Wunsch 1970 *J Mol Biol* 48: 443-453. Using GCG GAP software with the following settings for nucleic acid sequence comparison: GAP creation penalty of 5.0 and GAP extension penalty of 0.3, the coding region of the analogous nucleic acid sequences referred to above exhibits a degree of identity preferably of at least 70%, 75%, 80%, 85%, 90%, 95%, 98%, or 99%, with the CDS (encoding) part of the DNA sequence shown in SEQ ID NO: 1, 3, 5, 7, 9, or 11.

The term "sequence identity" refers to the degree to which two polynucleotide or polypeptide sequences are identical on a residue-by-residue basis over a particular region of comparison. The term "percentage of sequence identity" is calculated by comparing two optimally aligned sequences over that region of comparison, determining the number of positions at which the identical nucleic acid base (*e.g.*, A, T, C, G, U, or I, in the case of nucleic acids) occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the region of comparison (*i.e.*, the window size), and multiplying the result by 100 to yield the percentage of sequence identity. The term "substantial identity" as used herein denotes a characteristic of a polynucleotide sequence, wherein the polynucleotide comprises a sequence that has at least 80 percent sequence identity, preferably at least 85 percent identity and often 90 to 95 percent sequence identity, more usually at least 99 percent sequence identity as compared to a reference sequence over a comparison region.

Chimeric and fusion proteins

The invention also provides for BLAA chimeric or fusion proteins. As used herein, a BLAA "chimeric protein" or "fusion protein" comprises a BLAA polypeptide operatively linked to a non-BLAA polypeptide. A "BLAA polypeptide" refers to a polypeptide having an amino acid sequence corresponding to BLAA, whereas a "non-BLAA polypeptide" refers to a polypeptide having an amino acid sequence corresponding to a protein that is not substantially homologous to the BLAA polypeptide, *e.g.*, a protein that is different from the BLAA polypeptide and that is derived from the same or a different organism. Within a BLAA fusion protein the BLAA polypeptide can correspond to all or a portion of a BLAA polypeptide. In one embodiment, a BLAA fusion protein comprises at least one biologically active portion of a BLAA protein. In another embodiment, a BLAA fusion protein comprises at least two biologically active portions of a BLAA polypeptide. In yet another embodiment, a BLAA fusion protein comprises at least three biologically active portions of a BLAA polypeptide. Within the fusion protein, the term "operatively linked" is intended to indicate that the BLAA polypeptide and the non-BLAA polypeptide are fused in-frame to each other. The non-BLAA polypeptide can be fused to the N-terminus or C-terminus of the BLAA polypeptide.

Such fusion proteins can be further utilized in screening assays for compounds which modulate BLAA activity (such assays are described in detail below).

In one embodiment, the fusion protein is a GST-BLAA fusion protein in which the BLAA sequences are fused to the C-terminus of the GST (i.e., glutathione S-transferase) sequences. Such fusion proteins can facilitate the purification of recombinant BLAA.

In another embodiment, the fusion protein is a BLAA polypeptide containing a heterologous signal sequence at its N-terminus. For example, the nature BLAA signal sequence (e.g., about amino acids 1 to 26 of SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6) can be removed and replaced with a signal sequence from another protein. In certain host cells (e.g., mammalian host cells), expression and/or secretion of BLAA can be increased through use of a heterologous signal sequence.

In yet another embodiment, the fusion protein is a BLAA-immunoglobulin fusion protein in which the BLAA sequences are fused to sequences derived from a member of the immunoglobulin protein family. The BLAA-immunoglobulin fusion proteins of the invention can be incorporated into pharmaceutical compositions and administered to a subject to inhibit an interaction between a BLAA ligand and a BLAA protein on the surface of a cell, to thereby suppress BLAA-mediated signal transduction *in vivo*. The BLAA-immunoglobulin fusion proteins can be used to affect the bioavailability of a BLAA cognate ligand. Inhibition of the BLAA ligand/BLAA interaction may be useful therapeutically for both the treatment of immune response-associated disorders. Moreover, the BLAA-immunoglobulin fusion proteins of the invention can be used as immunogens to produce anti-BLAA antibodies in a subject, to purify BLAA ligands, and in screening assays to identify molecules that inhibit the interaction of BLAA with a BLAA ligand.

A BLAA chimeric or fusion protein of the invention can be produced by standard recombinant DNA techniques. For example, DNA fragments coding for the different polypeptide sequences are ligated together in-frame in accordance with conventional techniques, e.g., by employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including

automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers that give rise to complementary overhangs between two consecutive gene fragments that can subsequently be annealed and reamplified to generate a chimeric gene sequence (see, for example, Ausubel *et al.* (eds.) CURRENT PROTOCOLS IN MOLECULAR BIOLOGY, John Wiley & Sons, 1992). Moreover, many expression vectors are commercially available that already encode a fusion moiety (*e.g.*, a GST polypeptide). A BLAA-encoding nucleic acid can be cloned into such an expression vector such that the fusion moiety is linked in-frame to the BLAA polypeptide.

BLAA agonists and antagonists

The present invention also pertains to variants of the BLAA polypeptides that function as either BLAA agonists (mimetics) or as BLAA antagonists. Variants of the BLAA polypeptide can be generated by mutagenesis, *e.g.*, discrete point mutation or truncation of the BLAA polypeptide. An agonist of the BLAA polypeptide can retain substantially the same, or a subset of, the biological activities of the naturally occurring form of the BLAA polypeptide. An antagonist of the BLAA polypeptide can inhibit one or more of the activities of the naturally occurring form of the BLAA polypeptide by, for example, competitively binding to a downstream or upstream member of a cellular signaling cascade which includes the BLAA polypeptide. Thus, specific biological effects can be elicited by treatment with a variant of limited function. In one embodiment, treatment of a subject with a variant having a subset of the biological activities of the naturally occurring form of the polypeptide has fewer side effects in a subject relative to treatment with the naturally occurring form of the BLAA polypeptides.

Variants of the BLAA polypeptide that function as either BLAA agonists (mimetics) or as BLAA antagonists can be identified by screening combinatorial libraries of mutants, *e.g.*, truncation mutants, of the BLAA polypeptide for BLAA polypeptide agonist or antagonist activity. In one embodiment, a variegated library of BLAA variants is generated by combinatorial mutagenesis at the nucleic acid level and is encoded by a variegated gene library. A variegated library of BLAA variants can be produced by, for example, enzymatically ligating a mixture of synthetic oligonucleotides into gene sequences such that a degenerate set of potential BLAA sequences is expressible as individual polypeptides, or

alternatively, as a set of larger fusion proteins (*e.g.*, for phage display) containing the set of BLAA sequences therein. There are a variety of methods which can be used to produce libraries of potential BLAA variants from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be performed in an automatic DNA synthesizer, and the synthetic gene then ligated into an appropriate expression vector. Use of a degenerate set of genes allows for the provision, in one mixture, of all of the sequences encoding the desired set of potential BLAA sequences. Methods for synthesizing degenerate oligonucleotides are known in the art (see, *e.g.*, Narang (1983) *Tetrahedron* 39:3; Itakura *et al.* (1984) *Annu Rev Biochem* 53:323; Itakura *et al.* (1984) *Science* 198:1056; Ike *et al.* (1983) *Nucl Acid Res* 11:477.

Polypeptide libraries

In addition, libraries of fragments of the BLAA protein coding sequence can be used to generate a variegated population of BLAA fragments for screening and subsequent selection of variants of a BLAA polypeptide. In one embodiment, a library of coding sequence fragments can be generated by treating a double stranded PCR fragment of a BLAA coding sequence with a nuclease under conditions wherein nicking occurs only about once per molecule, denaturing the double stranded DNA, renaturing the DNA to form double stranded DNA that can include sense/antisense pairs from different nicked products, removing single stranded portions from reformed duplexes by treatment with S1 nuclease, and ligating the resulting fragment library into an expression vector. By this method, an expression library can be derived which encodes N-terminal and internal fragments of various sizes of a BLAA polypeptide.

Several techniques are known in the art for screening gene products of combinatorial libraries made by point mutations or truncation, and for screening cDNA libraries for gene products having a selected property. Such techniques are adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of a BLAA polypeptide. The most widely used techniques, which are amenable to high throughput analysis, for screening large gene libraries typically include cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates

isolation of the vector encoding the gene whose product was detected. Recursive ensemble mutagenesis (REM), a new technique that enhances the frequency of functional mutants in the libraries, can be used in combination with the screening assays to identify BLAA variants (Arkin and Yourvan (1992) *PNAS* 89:7811-7815; Delgrave *et al.* (1993) *Protein Engineering* 6:327-331).

Anti-BLAA antibodies

The invention encompasses antibodies and antibody fragments, such as F_{ab} or $(F_{ab})_2$, that bind immunospecifically to any of the polypeptides of the invention.

An isolated BLAA polypeptide, or a portion or fragment thereof, can be used as an immunogen to generate antibodies that bind BLAA using standard techniques for polyclonal and monoclonal antibody preparation. The full-length BLAA polypeptide can be used or, alternatively, the invention provides antigenic peptide fragments of BLAA for use as immunogens. The antigenic peptide of BLAA comprises at least 4 amino acid residues of the amino acid sequence shown in SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6 and encompasses an epitope of BLAA such that an antibody raised against the peptide forms a specific immune complex with BLAA. Preferably, the antigenic peptide comprises at least 6, 8, 10, 15, 20, or 30 amino acid residues. Longer antigenic peptides are sometimes preferable over shorter antigenic peptides, depending on use and according to methods well known to someone skilled in the art.

In certain embodiments of the invention, at least one epitope encompassed by the antigenic peptide is a region of BLAA that is located on the surface of the polypeptide, *e.g.*, a hydrophilic region. A hydrophobicity analysis of the human BLAA polypeptide sequence of SEQ ID NO:6, shown in FIG. 5, indicates that the hydrophilic regions include from about amino acid 75 to about amino acid 110; from about amino acid 150 to about amino acid 200; from about amino acid 225 to about amino acid 250; from about amino acid 290 to about amino acid 310; from about amino acid 380 to about amino acid 420; and from about amino acid 440 to about amino acid 534. These regions are likely to encode surface residues useful for targeting antibody production. As a means for targeting antibody production, hydropathy plots showing regions of hydrophilicity and hydrophobicity may be generated by any method well known in the art, including, for example, the Kyte Doolittle or the Hopp Woods methods,

either with or without Fourier transformation. See, *e.g.*, Hopp and Woods, 1981, *Proc. Nat. Acad. Sci. USA* 78: 3824-3828; Kyte and Doolittle 1982, *J. Mol. Biol.* 157: 105-142, each reference is incorporated herein by reference in their entirety.

As disclosed herein, BLAA polypeptide sequence of SEQ ID NO: 2, SEQ ID NO: 4, or
5 SEQ ID NO: 6, or derivatives, fragments, analogs or homologs thereof, may be utilized as immunogens in the generation of antibodies that immunospecifically-bind these protein components. The term "antibody" as used herein refers to immunoglobulin molecules and immunologically active portions of immunoglobulin molecules, *i.e.*, molecules that contain an antigen binding site that specifically binds (immunoreacts with) an antigen, such as BLAA.
10 Such antibodies include, but are not limited to, polyclonal, monoclonal, chimeric, single chain, F_{ab} and $F_{(ab')_2}$ fragments, and an F_{ab} expression library. In a specific embodiment, antibodies to human BLAA polypeptides are disclosed. Various procedures known within the art may be used for the production of polyclonal or monoclonal antibodies to a BLAA polypeptide sequence of SEQ ID NO: 2, SEQ ID NO: 4, or SEQ ID NO: 6, or derivative, fragment, analog
15 or homolog thereof. Some of these proteins are discussed below.

For the production of polyclonal antibodies, various suitable host animals (*e.g.*, rabbit, goat, mouse or other mammal) may be immunized by injection with the native polypeptide, or a synthetic variant thereof, or a derivative of the foregoing. An appropriate immunogenic preparation can contain, for example, recombinantly expressed BLAA polypeptide or a
20 chemically synthesized BLAA polypeptide. The preparation can further include an adjuvant. Various adjuvants used to increase the immunological response include, but are not limited to, Freund's (complete and incomplete), mineral gels (*e.g.*, aluminum hydroxide), surface active substances (*e.g.*, lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, dinitrophenol, etc.), human adjuvants such as *Bacille Calmette-Guerin* and *Corynebacterium*
25 *parvum*, or similar immunostimulatory agents. If desired, the antibody molecules directed against BLAA can be isolated from the mammal (*e.g.*, from the blood) and further purified by well known techniques, such as protein A chromatography to obtain the IgG fraction.

The term "monoclonal antibody" or "monoclonal antibody composition", as used herein, refers to a population of antibody molecules that contain only one species of an antigen
30 binding site capable of immunoreacting with a particular epitope of BLAA. A monoclonal

antibody composition thus typically displays a single binding affinity for a particular BLAA polypeptide with which it immunoreacts. For preparation of monoclonal antibodies directed towards a particular BLAA polypeptide, or derivatives, fragments, analogs or homologs thereof, any technique that provides for the production of antibody molecules by continuous cell line culture may be utilized. Such techniques include, but are not limited to, the hybridoma technique (see Kohler & Milstein, 1975 *Nature* 256: 495-497); the trioma technique; the human B-cell hybridoma technique (see Kozbor, *et al.*, 1983 *Immunol Today* 4: 72) and the EBV hybridoma technique to produce human monoclonal antibodies (see Cole, *et al.*, 1985 In: MONOCLONAL ANTIBODIES AND CANCER THERAPY, Alan R. Liss, Inc., pp. 77-96). Human monoclonal antibodies may be utilized in the practice of the present invention and may be produced by using human hybridomas (see Cote, *et al.*, 1983. *Proc Natl Acad Sci USA* 80: 2026-2030) or by transforming human B-cells with Epstein Barr Virus *in vitro* (see Cole, *et al.*, 1985 In: MONOCLONAL ANTIBODIES AND CANCER THERAPY, Alan R. Liss, Inc., pp. 77-96). Each of the above citations are incorporated herein by reference in their entirety.

According to the invention, techniques can be adapted for the production of single-chain antibodies specific to a BLAA polypeptide (see *e.g.*, U.S. Patent No. 4,946,778). In addition, methodologies can be adapted for the construction of F_{ab} expression libraries (see *e.g.*, Huse, *et al.*, 1989 *Science* 246: 1275-1281) to allow rapid and effective identification of monoclonal F_{ab} fragments with the desired specificity for a BLAA polypeptide or derivatives, fragments, analogs or homologs thereof. Non-human antibodies can be "humanized" by techniques well known in the art. See *e.g.*, U.S. Patent No. 5,225,539. Antibody fragments that contain the idiotypes to a BLAA polypeptide may be produced by techniques known in the art including, but not limited to: (i) an $F_{(ab)_2}$ fragment produced by pepsin digestion of an antibody molecule; (ii) an F_{ab} fragment generated by reducing the disulfide bridges of an $F_{(ab)_2}$ fragment; (iii) an F_{ab} fragment generated by the treatment of the antibody molecule with papain and a reducing agent and (iv) F_c fragments.

Additionally, recombinant anti-BLAA antibodies, such as chimeric and humanized monoclonal antibodies, comprising both human and non-human portions, which can be made using standard recombinant DNA techniques, are within the scope of the invention. Such chimeric and humanized monoclonal antibodies can be produced by recombinant DNA

techniques known in the art, for example using methods described in International Application No. PCT/US86/02269; European Patent Application No. 184,187; European Patent Application No. 171,496; European Patent Application No. 173,494; PCT International Publication No. WO 86/01533; U.S. Pat. No. 4,816,567; U.S. Pat. No. 5,225,539; European Patent Application No. 125,023; Better *et al.* (1988) *Science* 240:1041-1043; Liu *et al.* (1987) *PNAS* 84:3439-3443; Liu *et al.* (1987) *J Immunol.* 139:3521-3526; Sun *et al.* (1987) *PNAS* 84:214-218; Nishimura *et al.* (1987) *Cancer Res* 47:999-1005; Wood *et al.* (1985) *Nature* 314:446-449; Shaw *et al.* (1988) *J Natl Cancer Inst* 80:1553-1559; Morrison (1985) *Science* 229:1202-1207; Oi *et al.* (1986) *BioTechniques* 4:214; Jones *et al.* (1986) *Nature* 321:552-525; Verhoeyan *et al.* (1988) *Science* 239:1534; and Beidler *et al.* (1988) *J Immunol* 141:4053-4060. Each of the above citations are incorporated herein by reference in their entirety.

In one embodiment, methodologies for the screening of antibodies that possess the desired specificity include, but are not limited to, enzyme-linked immunosorbent assay (ELISA) and other immunologically-mediated techniques known within the art. In a specific embodiment, selection of antibodies that are specific to a particular domain of a BLAA polypeptide is facilitated by generation of hybridomas that bind to the fragment of a BLAA polypeptide possessing such a domain. Antibodies that are specific for an Ig-like domain within a BLAA polypeptide, or derivatives, fragments, analogs or homologs thereof, are also provided herein.

Anti-BLAA antibodies may be used in methods known within the art relating to the localization and/or quantitation of a BLAA polypeptide (*e.g.*, for use in measuring levels of the BLAA polypeptide within appropriate physiological samples, for use in diagnostic methods, for use in imaging the polypeptide, and the like). In a given embodiment, antibodies for BLAA polypeptides, or derivatives, fragments, analogs or homologs thereof, that contain the antibody derived binding domain, are utilized as pharmacologically-active compounds [hereinafter "therapeutics"].

An anti-BLAA antibody (*e.g.*, monoclonal antibody) can be used to isolate BLAA by standard techniques, such as affinity chromatography or immunoprecipitation. An anti-BLAA antibody can facilitate the purification of natural BLAA from cells and of recombinantly

produced BLAA expressed in host cells. Moreover, an anti-BLAA antibody can be used to detect BLAA polypeptide (*e.g.*, in a cellular lysate or cell supernatant) in order to evaluate the abundance and pattern of expression of the BLAA polypeptide. Anti-BLAA antibodies can be used diagnostically to monitor protein levels in tissue as part of a clinical testing procedure, *e.g.*, to, for example, determine the efficacy of a given treatment regimen. Detection can be facilitated by coupling (*i.e.*, physically linking) the antibody to a detectable substance. Examples of detectable substances include various enzymes, prosthetic groups, fluorescent materials, luminescent materials, bioluminescent materials, and radioactive materials. Examples of suitable enzymes include horseradish peroxidase, alkaline phosphatase, β -galactosidase, or acetylcholinesterase; examples of suitable prosthetic group complexes include streptavidin/biotin and avidin/biotin; examples of suitable fluorescent materials include umbelliferone, fluorescein, fluorescein isothiocyanate, rhodamine, dichlorotriazinylamine fluorescein, dansyl chloride or phycoerythrin; an example of a luminescent material includes luminol; examples of bioluminescent materials include luciferase, luciferin, and aequorin, and examples of suitable radioactive material include ^{125}I , ^{131}I , ^{35}S or ^3H .

BLAA Recombinant Expression Vectors and Host Cells

Another aspect of the invention pertains to vectors, preferably expression vectors, containing a nucleic acid encoding BLAA polypeptide, or derivatives, fragments, analogs or homologs thereof. As used herein, the term "vector" refers to a nucleic acid molecule capable of transporting another nucleic acid to which it has been linked. One type of vector is a "plasmid", which refers to a circular double stranded DNA loop into which additional DNA segments can be ligated. Another type of vector is a viral vector, wherein additional DNA segments can be ligated into the viral genome. Certain vectors are capable of autonomous replication in a host cell into which they are introduced (*e.g.*, bacterial vectors having a bacterial origin of replication and episomal mammalian vectors). Other vectors (*e.g.*, non-episomal mammalian vectors) are integrated into the genome of a host cell upon introduction into the host cell, and thereby are replicated along with the host genome. Moreover, certain vectors are capable of directing the expression of genes to which they are operatively linked. Such vectors are referred to herein as "expression vectors". In general,

expression vectors of utility in recombinant DNA techniques are often in the form of plasmids. In the present specification, "plasmid" and "vector" can be used interchangeably as the plasmid is the most commonly used form of vector. However, the invention is intended to include such other forms of expression vectors, such as viral vectors (*e.g.*, replication defective retroviruses, adenoviruses and adeno-associated viruses), which serve equivalent functions.

The recombinant expression vectors of the invention comprise a nucleic acid of the invention in a form suitable for expression of the nucleic acid in a host cell, which means that the recombinant expression vectors include one or more regulatory sequences, selected on the basis of the host cells to be used for expression, that is operatively linked to the nucleic acid sequence to be expressed. Within a recombinant expression vector, "operably linked" is intended to mean that the nucleotide sequence of interest is linked to the regulatory sequence(s) in a manner that allows for expression of the nucleotide sequence (*e.g.*, in an *in vitro* transcription/translation system or in a host cell when the vector is introduced into the host cell). The term "regulatory sequence" is intended to include promoters, enhancers and other expression control elements (*e.g.*, polyadenylation signals). Such regulatory sequences are described, for example, in Goeddel; GENE EXPRESSION TECHNOLOGY: METHODS IN ENZYMOLOGY 185, Academic Press, San Diego, Calif. (1990). Regulatory sequences include those that direct constitutive expression of a nucleotide sequence in many types of host cell and those that direct expression of the nucleotide sequence only in certain host cells (*e.g.*, tissue-specific regulatory sequences). It will be appreciated by those skilled in the art that the design of the expression vector can depend on such factors as the choice of the host cell to be transformed, the level of expression of protein desired, etc. The expression vectors of the invention can be introduced into host cells to thereby produce proteins or peptides, including fusion proteins or peptides, encoded by nucleic acids as described herein (*e.g.*, BLAA polypeptides, mutant forms of BLAA, fusion proteins, etc.).

The recombinant expression vectors of the invention can be designed for expression of BLAA in prokaryotic or eukaryotic cells. For example, BLAA can be expressed in bacterial cells such as *E. coli*, insect cells (using baculovirus expression vectors) yeast cells or mammalian cells. Suitable host cells are discussed further in Goeddel, GENE EXPRESSION TECHNOLOGY: METHODS IN ENZYMOLOGY 185, Academic Press, San Diego, Calif. (1990).

Alternatively, the recombinant expression vector can be transcribed and translated *in vitro*, for example using T7 promoter regulatory sequences and T7 polymerase.

Expression of proteins in prokaryotes is most often carried out in *E. coli* with vectors containing constitutive or inducible promoters directing the expression of either fusion or non-fusion proteins. Fusion vectors add a number of amino acids to a protein encoded therein, usually to the amino terminus of the recombinant protein. Such fusion vectors typically serve three purposes: (1) to increase expression of recombinant protein; (2) to increase the solubility of the recombinant protein; and (3) to aid in the purification of the recombinant protein by acting as a ligand in affinity purification. Often, in fusion expression vectors, a proteolytic cleavage site is introduced at the junction of the fusion moiety and the recombinant protein to enable separation of the recombinant protein from the fusion moiety subsequent to purification of the fusion protein. Such enzymes, and their cognate recognition sequences, include Factor Xa, thrombin and enterokinase. Typical fusion expression vectors include pGEX (Pharmacia Biotech Inc; Smith and Johnson (1988) *Gene* 67:31-40), pMAL (New England Biolabs, Beverly, Mass.) and pRIT5 (Pharmacia, Piscataway, N.J.) that fuse glutathione S-transferase (GST), maltose E binding protein, or protein A, respectively, to the target recombinant protein.

Examples of suitable inducible non-fusion *E. coli* expression vectors include pTrc (Amrann *et al.*, (1988) *Gene* 69:301-315) and pET 11d (Studier *et al.*, GENE EXPRESSION TECHNOLOGY: METHODS IN ENZYMOLOGY 185, Academic Press, San Diego, Calif. (1990) 60-89).

One strategy to maximize recombinant protein expression in *E. coli* is to express the protein in a host bacteria with an impaired capacity to proteolytically cleave the recombinant protein. See, Gottesman, GENE EXPRESSION TECHNOLOGY: METHODS IN ENZYMOLOGY 185, Academic Press, San Diego, Calif. (1990) 119-128. Another strategy is to alter the nucleic acid sequence of the nucleic acid to be inserted into an expression vector so that the individual codons for each amino acid are those preferentially utilized in *E. coli* (Wada *et al.*, (1992) *Nucleic Acids Res.* 20:2111-2118). Such alteration of nucleic acid sequences of the invention can be carried out by standard DNA synthesis techniques.

In another embodiment, the BLAA expression vector is a yeast expression vector. Examples of vectors for expression in yeast *S. cerevisiae* include pYepSec1 (Baldari, *et al.*,

(1987) *EMBO J* 6:229-234), pMFa (Kurjan and Herskowitz, (1982) *Cell* 30:933-943), pJRY88 (Schultz *et al.*, (1987) *Gene* 54:113-123), pYES2 (Invitrogen Corporation, San Diego, Calif.), and picZ (Invitrogen Corp, San Diego, Calif.).

Alternatively, BLAA can be expressed in insect cells using baculovirus expression
5 vectors. Baculovirus vectors available for expression of proteins in cultured insect cells (*e.g.*, SF9 cells) include the pAc series (Smith *et al.* (1983) *Mol Cell Biol* 3:2156-2165) and the pVL series (Lucklow and Summers (1989) *Virology* 170:31-39).

In yet another embodiment, a nucleic acid of the invention is expressed in mammalian cells using a mammalian expression vector. Examples of mammalian expression vectors
10 include pCDM8 (Seed (1987) *Nature* 329:840) and pMT2PC (Kaufman *et al.* (1987) *EMBO J* 6: 187-195). When used in mammalian cells, the expression vector's control functions are often provided by viral regulatory elements. For example, commonly used promoters are derived from polyoma, Adenovirus 2, cytomegalovirus and Simian Virus 40. For other suitable expression systems for both prokaryotic and eukaryotic cells. See, *e.g.*, Chapters 16
15 and 17 of Sambrook *et al.*, MOLECULAR CLONING: A LABORATORY MANUAL. 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989.

In another embodiment, the recombinant mammalian expression vector is capable of directing expression of the nucleic acid preferentially in a particular cell type (*e.g.*,
20 tissue-specific regulatory elements are used to express the nucleic acid). Tissue-specific regulatory elements are known in the art. Non-limiting examples of suitable tissue-specific promoters include the albumin promoter (liver-specific; Pinkert *et al.* (1987) *Genes Dev* 1:268-277), lymphoid-specific promoters (Calame and Eaton (1988) *Adv Immunol* 43:235-275), in particular promoters of T cell receptors (Winoto and Baltimore (1989) *EMBO J* 8:729-733) and immunoglobulins (Banerji *et al.* (1983) *Cell* 33:729-740; Queen and Baltimore (1983) *Cell* 33:741-748), neuron-specific promoters (*e.g.*, the neurofilament promoter; Byrne and Ruddle (1989) *PNAS* 86:5473-5477), pancreas-specific promoters (Edlund *et al.* (1985) *Science* 230:912-916), and mammary gland-specific promoters (*e.g.*, milk whey promoter; U.S. Pat. No. 4,873,316 and European Application Publication No.
25 264,166). Developmentally-regulated promoters are also encompassed, *e.g.*, the murine hox
30 264,166).

promoters (Kessel and Gruss (1990) *Science* 249:374-379) and the α -fetoprotein promoter (Campes and Tilghman (1989) *Genes Dev* 3:537-546).

The invention further provides a recombinant expression vector comprising a DNA molecule of the invention cloned into the expression vector in an antisense orientation. That
5 is, the DNA molecule is operatively linked to a regulatory sequence in a manner that allows for expression (by transcription of the DNA molecule) of an RNA molecule that is antisense to BLAA mRNA. Regulatory sequences operatively linked to a nucleic acid cloned in the antisense orientation can be chosen that direct the continuous expression of the antisense RNA molecule in a variety of cell types, for instance viral promoters and/or enhancers, or regulatory
10 sequences can be chosen that direct constitutive, tissue specific or cell type specific expression of antisense RNA. The antisense expression vector can be in the form of a recombinant plasmid, phagemid or attenuated virus in which antisense nucleic acids are produced under the control of a high efficiency regulatory region, the activity of which can be determined by the cell type into which the vector is introduced. For a discussion of the regulation of gene
15 expression using antisense genes see Weintraub *et al.*, "Antisense RNA as a molecular tool for genetic analysis," Reviews--Trends in Genetics, Vol. 1(1) 1986.

Another aspect of the invention pertains to host cells into which a recombinant expression vector of the invention has been introduced. The terms "host cell" and "recombinant host cell" are used interchangeably herein. It is understood that such terms refer
20 not only to the particular subject cell but to the progeny or potential progeny of such a cell. Because certain modifications may occur in succeeding generations due to either mutation or environmental influences, such progeny may not, in fact, be identical to the parent cell, but are still included within the scope of the term as used herein.

A host cell can be any prokaryotic or eukaryotic cell. For example, BLAA polypeptide
25 can be expressed in bacterial cells such as *E. coli*, insect cells, yeast or mammalian cells (such as Chinese hamster ovary cells (CHO) or COS cells). Other suitable host cells are known to those skilled in the art.

Vector DNA can be introduced into prokaryotic or eukaryotic cells via conventional transformation or transfection techniques. As used herein, the terms "transformation" and
30 "transfection" are intended to refer to a variety of art-recognized techniques for introducing

foreign nucleic acid (*e.g.*, DNA) into a host cell, including calcium phosphate or calcium chloride co-precipitation, DEAE-dextran-mediated transfection, lipofection, or electroporation. Suitable methods for transforming or transfecting host cells can be found in Sambrook, *et al.* (MOLECULAR CLONING: A LABORATORY MANUAL. 2nd ed., Cold Spring Harbor Laboratory, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y., 1989), and other laboratory manuals.

For stable transfection of mammalian cells, it is known that, depending upon the expression vector and transfection technique used, only a small fraction of cells may integrate the foreign DNA into their genome. In order to identify and select these integrants, a gene that encodes a selectable marker (*e.g.*, resistance to antibiotics) is generally introduced into the host cells along with the gene of interest. Various selectable markers include those that confer resistance to drugs, such as G418, hygromycin and methotrexate. Nucleic acid encoding a selectable marker can be introduced into a host cell on the same vector as that encoding BLAA or can be introduced on a separate vector. Cells stably transfected with the introduced nucleic acid can be identified by drug selection (*e.g.*, cells that have incorporated the selectable marker gene will survive, while the other cells die).

A host cell of the invention, such as a prokaryotic or eukaryotic host cell in culture, can be used to produce (*i.e.*, express) BLAA polypeptide. Accordingly, the invention further provides methods for producing BLAA polypeptide using the host cells of the invention. In one embodiment, the method comprises culturing the host cell of invention (into which a recombinant expression vector encoding BLAA has been introduced) in a suitable medium such that BLAA polypeptide is produced. In another embodiment, the method further comprises isolating BLAA from the medium or the host cell.

Transgenic animals

The host cells of the invention can also be used to produce nonhuman transgenic animals. For example, in one embodiment, a host cell of the invention is a fertilized oocyte or an embryonic stem cell into which BLAA-coding sequences have been introduced. Such host cells can then be used to create non-human transgenic animals in which exogenous BLAA sequences have been introduced into their genome or homologous recombinant animals in which endogenous BLAA sequences have been altered. Such animals are useful for studying

the function and/or activity of BLAA and for identifying and/or evaluating modulators of BLAA activity. As used herein, a "transgenic animal" is a non-human animal, preferably a mammal, more preferably a rodent such as a rat or mouse, in which one or more of the cells of the animal includes a transgene. Other examples of transgenic animals include non-human
5 primates, sheep, dogs, cows, goats, chickens, amphibians, etc. A transgene is exogenous DNA that is integrated into the genome of a cell from which a transgenic animal develops and that remains in the genome of the mature animal, thereby directing the expression of an encoded gene product in one or more cell types or tissues of the transgenic animal. As used herein, a "homologous recombinant animal" is a non-human animal, preferably a mammal, more
10 preferably a mouse, in which an endogenous BLAA gene has been altered by homologous recombination between the endogenous gene and an exogenous DNA molecule introduced into a cell of the animal, *e.g.*, an embryonic cell of the animal, prior to development of the animal.

A transgenic animal of the invention can be created by introducing BLAA-encoding nucleic acid into the male pronuclei of a fertilized oocyte, *e.g.*, by microinjection, retroviral
15 infection, and allowing the oocyte to develop in a pseudopregnant female foster animal. The human BLAA cDNA sequence of SEQ ID NO: 1, 3, 5, 7, 9, or 11, can be introduced as a transgene into the genome of a non-human animal. Alternatively, a nonhuman homologue of the human BLAA gene, such as a mouse BLAA gene, can be isolated based on hybridization to the human BLAA cDNA (described further above) and used as a transgene. Intronic
20 sequences and polyadenylation signals can also be included in the transgene to increase the efficiency of expression of the transgene. A tissue-specific regulatory sequence(s) can be operably linked to the BLAA transgene to direct expression of BLAA polypeptide to particular cells. Methods for generating transgenic animals via embryo manipulation and microinjection, particularly animals such as mice, have become conventional in the art and are described, for
25 example, in U.S. Pat. Nos. 4,736,866; 4,870,009; and 4,873,191; and Hogan 1986, In: MANIPULATING THE MOUSE EMBRYO, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, N.Y. Similar methods are used for production of other transgenic animals. A transgenic founder animal can be identified based upon the presence of the BLAA transgene in its genome and/or expression of BLAA mRNA in tissues or cells of the animals. A transgenic
30 founder animal can then be used to breed additional animals carrying the transgene.

Moreover, transgenic animals carrying a transgene encoding BLAA can further be bred to other transgenic animals carrying other transgenes.

To create a homologous recombinant animal, a vector is prepared which contains at least a portion of a BLAA gene into which a deletion, addition or substitution has been introduced to thereby alter, *e.g.*, functionally disrupt, the BLAA gene. The BLAA gene can be a human gene (*e.g.*, the cDNA of SEQ ID NO: 1, 3, 5, 7, 9, or 11), but more preferably, is a non-human homologue of a human BLAA gene. For example, a mouse homologue of human BLAA gene of SEQ ID NO: 1, 3, 5, 7, 9, or 11 can be used to construct a homologous recombination vector suitable for altering an endogenous BLAA gene in the mouse genome. In one embodiment, the vector is designed such that, upon homologous recombination, the endogenous BLAA gene is functionally disrupted (*i.e.*, no longer encodes a functional protein; also referred to as a "knock out" vector).

Alternatively, the vector can be designed such that, upon homologous recombination, the endogenous BLAA gene is mutated or otherwise altered but still encodes functional protein (*e.g.*, the upstream regulatory region can be altered to thereby alter the expression of the endogenous BLAA polypeptide). In the homologous recombination vector, the altered portion of the BLAA gene is flanked at its 5' and 3' ends by additional nucleic acid of the BLAA gene to allow for homologous recombination to occur between the exogenous BLAA gene carried by the vector and an endogenous BLAA gene in an embryonic stem cell. The additional flanking BLAA nucleic acid is of sufficient length for successful homologous recombination with the endogenous gene. Typically, several kilobases of flanking DNA (both at the 5' and 3' ends) are included in the vector. See *e.g.*, Thomas *et al.* (1987) *Cell* 51:503 for a description of homologous recombination vectors. The vector is introduced into an embryonic stem cell line (*e.g.*, by electroporation) and cells in which the introduced BLAA gene has homologously recombined with the endogenous BLAA gene are selected (see *e.g.*, Li *et al.* (1992) *Cell* 69:915).

The selected cells are then injected into a blastocyst of an animal (*e.g.*, a mouse) to form aggregation chimeras. See *e.g.*, Bradley 1987, In: TERATOCARCINOMAS AND EMBRYONIC STEM CELLS: A PRACTICAL APPROACH, Robertson, ed. IRL, Oxford, pp. 113-152. A chimeric embryo can then be implanted into a suitable pseudopregnant female foster animal and the

embryo brought to term. Progeny harboring the homologously recombined DNA in their germ cells can be used to breed animals in which all cells of the animal contain the homologously recombined DNA by germline transmission of the transgene. Methods for constructing homologous recombination vectors and homologous recombinant animals are described further in Bradley (1991) *Curr Opin Biotechnol* 2:823-829; PCT International Publication Nos.: WO 90/11354; WO 91/01140; WO 92/0968; and WO 93/04169.

In another embodiment, transgenic non-humans animals can be produced that contain selected systems that allow for regulated expression of the transgene. One example of such a system is the cre/loxP recombinase system of bacteriophage P1. For a description of the cre/loxP recombinase system, see, *e.g.*, Lakso *et al.* (1992) *PNAS* 89:6232-6236. Another example of a recombinase system is the FLP recombinase system of *Saccharomyces cerevisiae* (O'Gorman *et al.* (1991) *Science* 251:1351-1355. If a cre/loxP recombinase system is used to regulate expression of the transgene, animals containing transgenes encoding both the Cre recombinase and a selected protein are required. Such animals can be provided through the construction of "double" transgenic animals, *e.g.*, by mating two transgenic animals, one containing a transgene encoding a selected protein and the other containing a transgene encoding a recombinase.

Clones of the non-human transgenic animals described herein can also be produced according to the methods described in Wilmut *et al.* (1997) *Nature* 385:810-813. In brief, a cell, *e.g.*, a somatic cell, from the transgenic animal can be isolated and induced to exit the growth cycle and enter G₀ phase. The quiescent cell can then be fused, *e.g.*, through the use of electrical pulses, to an enucleated oocyte from an animal of the same species from which the quiescent cell is isolated. The reconstructed oocyte is then cultured such that it develops to morula or blastocyte and then transferred to pseudopregnant female foster animal. The offspring borne of this female foster animal will be a clone of the animal from which the cell, *e.g.*, the somatic cell, is isolated.

Pharmaceutical Compositions

The BLAA nucleic acid molecules, BLAA polypeptides, and anti-BLAA antibodies (also referred to herein as "active compounds") of the invention, and derivatives, fragments, analogs and homologs thereof, can be incorporated into pharmaceutical compositions suitable

for administration. Such compositions typically comprise the nucleic acid molecule, protein, or antibody and a pharmaceutically acceptable carrier. As used herein, "pharmaceutically acceptable carrier" is intended to include any and all solvents, dispersion media, coatings, antibacterial and antifungal agents, isotonic and absorption delaying agents, and the like, compatible with pharmaceutical administration. Suitable carriers are described in the most recent edition of Remington's Pharmaceutical Sciences, a standard reference text in the field, which is incorporated herein by reference. Preferred examples of such carriers or diluents include, but are not limited to, water, saline, finger's solutions, dextrose solution, and 5% human serum albumin. Liposomes and non-aqueous vehicles such as fixed oils may also be used. The use of such media and agents for pharmaceutically active substances is well known in the art. Except insofar as any conventional media or agent is incompatible with the active compound, use thereof in the compositions is contemplated. Supplementary active compounds can also be incorporated into the compositions.

A pharmaceutical composition of the invention is formulated to be compatible with its intended route of administration. Examples of routes of administration include parenteral, *e.g.*, intravenous, intradermal, subcutaneous, oral (*e.g.*, inhalation), transdermal (topical), transmucosal, and rectal administration. Solutions or suspensions used for parenteral, intradermal, or subcutaneous application can include the following components: a sterile diluent such as water for injection, saline solution, fixed oils, polyethylene glycols, glycerine, propylene glycol or other synthetic solvents; antibacterial agents such as benzyl alcohol or methyl parabens; antioxidants such as ascorbic acid or sodium bisulfite; chelating agents such as ethylenediaminetetraacetic acid; buffers such as acetates, citrates or phosphates, and agents for the adjustment of tonicity such as sodium chloride or dextrose. The pH can be adjusted with acids or bases, such as hydrochloric acid or sodium hydroxide. The parenteral preparation can be enclosed in ampoules, disposable syringes or multiple dose vials made of glass or plastic.

Pharmaceutical compositions suitable for injectable use include sterile aqueous solutions (where water soluble) or dispersions and sterile powders for the extemporaneous preparation of sterile injectable solutions or dispersion. For intravenous administration, suitable carriers include physiological saline, bacteriostatic water, Cremophor EL™ (BASF,

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Parsippany, N.J.) or phosphate buffered saline (PBS). In all cases, the composition must be sterile and should be fluid to the extent that easy syringeability exists. It must be stable under the conditions of manufacture and storage and must be preserved against the contaminating action of microorganisms such as bacteria and fungi. The carrier can be a solvent or
5 dispersion medium containing, for example, water, ethanol, polyol (for example, glycerol, propylene glycol, and liquid polyethylene glycol, and the like), and suitable mixtures thereof. The proper fluidity can be maintained, for example, by the use of a coating such as lecithin, by the maintenance of the required particle size in the case of dispersion and by the use of surfactants. Prevention of the action of microorganisms can be achieved by various
10 antibacterial and antifungal agents, for example, parabens, chlorobutanol, phenol, ascorbic acid, thimerosal, and the like. In many cases, it will be preferable to include isotonic agents, for example, sugars, polyalcohols such as manitol, sorbitol, sodium chloride in the composition. Prolonged absorption of the injectable compositions can be brought about by including in the composition an agent which delays absorption, for example, aluminum
15 monostearate and gelatin.

Sterile injectable solutions can be prepared by incorporating the active compound (e.g., a BLAA polypeptide or anti-BLAA antibody) in the required amount in an appropriate solvent with one or a combination of ingredients enumerated above, as required, followed by filtered sterilization. Generally, dispersions are prepared by incorporating the active compound into a
20 sterile vehicle that contains a basic dispersion medium and the required other ingredients from those enumerated above. In the case of sterile powders for the preparation of sterile injectable solutions, methods of preparation are vacuum drying and freeze-drying that yields a powder of the active ingredient plus any additional desired ingredient from a previously sterile-filtered solution thereof.

25 Oral compositions generally include an inert diluent or an edible carrier. They can be enclosed in gelatin capsules or compressed into tablets. For the purpose of oral therapeutic administration, the active compound can be incorporated with excipients and used in the form of tablets, troches, or capsules. Oral compositions can also be prepared using a fluid carrier for use as a mouthwash, wherein the compound in the fluid carrier is applied orally and
30 swished and expectorated or swallowed. Pharmaceutically compatible binding agents, and/or

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adjuvant materials can be included as part of the composition. The tablets, pills, capsules, troches and the like can contain any of the following ingredients, or compounds of a similar nature: a binder such as microcrystalline cellulose, gum tragacanth or gelatin; an excipient such as starch or lactose, a disintegrating agent such as alginic acid, Primogel, or corn starch; a
5 lubricant such as magnesium stearate or Sterotes; a glidant such as colloidal silicon dioxide; a sweetening agent such as sucrose or saccharin; or a flavoring agent such as peppermint, methyl salicylate, or orange flavoring.

For administration by inhalation, the compounds are delivered in the form of an aerosol spray from pressured container or dispenser which contains a suitable propellant, *e.g.*, a gas
10 such as carbon dioxide, or a nebulizer.

Systemic administration can also be by transmucosal or transdermal means. For transmucosal or transdermal administration, penetrants appropriate to the barrier to be permeated are used in the formulation. Such penetrants are generally known in the art, and include, for example, for transmucosal administration, detergents, bile salts, and fusidic acid
15 derivatives. Transmucosal administration can be accomplished through the use of nasal sprays or suppositories. For transdermal administration, the active compounds are formulated into ointments, salves, gels, or creams as generally known in the art.

The compounds can also be prepared in the form of suppositories (*e.g.*, with conventional suppository bases such as cocoa butter and other glycerides) or retention enemas for rectal
20 delivery.

In one embodiment, the active compounds are prepared with carriers that will protect the compound against rapid elimination from the body, such as a controlled release formulation, including implants and microencapsulated delivery systems. Biodegradable, biocompatible polymers can be used, such as ethylene vinyl acetate, polyanhydrides,
25 polyglycolic acid, collagen, polyorthoesters, and polylactic acid. Methods for preparation of such formulations will be apparent to those skilled in the art. The materials can also be obtained commercially from Alza Corporation and Nova Pharmaceuticals, Inc. Liposomal suspensions (including liposomes targeted to infected cells with monoclonal antibodies to viral antigens) can also be used as pharmaceutically acceptable carriers. These can be prepared

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according to methods known to those skilled in the art, for example, as described in U.S. Pat. No. 4,522,811.

It is especially advantageous to formulate oral or parenteral compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used
5 herein refers to physically discrete units suited as unitary dosages for the subject to be treated; each unit containing a predetermined quantity of active compound calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. The specification for the dosage unit forms of the invention are dictated by and directly dependent on the unique characteristics of the active compound and the particular therapeutic effect to be
10 achieved, and the limitations inherent in the art of compounding such an active compound for the treatment of individuals.

The nucleic acid molecules of the invention can be inserted into vectors and used as gene therapy vectors. Gene therapy vectors can be delivered to a subject by, for example, intravenous injection, local administration (see U.S. Pat. No. 5,328,470) or by stereotactic
15 injection (see *e.g.*, Chen *et al.* (1994) *PNAS* 91:3054-3057). The pharmaceutical preparation of the gene therapy vector can include the gene therapy vector in an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery vector can be produced intact from recombinant cells, *e.g.*, retroviral vectors, the pharmaceutical preparation can include one or more cells that produce
20 the gene delivery system.

The pharmaceutical compositions can be included in a container, pack, or dispenser together with instructions for administration.

Uses and Methods of the Invention

The immunoglobulin superfamily members include multifunctional secreted and
25 membrane-bound proteins that modulate a number of functions. B7 molecules known in the prior art are located on the cell surface. (See Selvakumar *et al.* 1992, *Immunogenetics* 36:175-181 and Linsley *et al.* 1994, *Protein Sc.* 3:1341-1343). Sequence analysis studies using programs known in the prior art (*e.g.*, PSORT) show that the polypeptides of the instant invention are most likely localized in the membrane of the endoplasmic reticulum. The

nucleic acid molecules, proteins, protein homologues, and antibodies described herein can be used in one or more of the following methods: (a) screening assays; (b) detection assays (*e.g.*, chromosomal mapping, tissue typing, forensic biology), (c) predictive medicine (*e.g.*, diagnostic assays, prognostic assays, monitoring clinical trials, and pharmacogenomics); and

5 (d) methods of treatment (*e.g.*, therapeutic and prophylactic). As described herein, in one embodiment, a BLAA polypeptide of the invention has the ability to bind to activated T lymphocytes and provide regulatory signals for T lymphocyte cell growth and activation. A BLAA polypeptide interacts with other cellular proteins and can thus be used to modulate immune response-associated protein activity. Such modulation may have an effect on the

10 regulation of cellular proliferation, the regulation of cellular differentiation, and/or the regulation of cell survival.

The isolated nucleic acid molecules of the invention can be used to express BLAA polypeptide (*e.g.*, via a recombinant expression vector in a host cell in gene therapy applications), to detect BLAA mRNA (*e.g.*, in a biological sample) or a genetic lesion in a

15 BLAA gene, and to modulate BLAA activity, as described further below. In addition, the BLAA polypeptides can be used to screen drugs or compounds that modulate the BLAA activity or expression as well as to treat disorders characterized by insufficient or excessive production of BLAA polypeptide or production of BLAA polypeptide forms that have decreased or aberrant activity compared to BLAA wild type polypeptide (*e.g.* infectious

20 diseases, cancers, autoimmune disorders, and complications associated with transplantation). In addition, the anti-BLAA antibodies of the invention can be used to detect and isolate BLAA polypeptides and modulate BLAA activity.

This invention further pertains to novel agents identified by the above described screening assays and uses thereof for treatments as described herein.

25 **Screening Assays**

The invention provides a method (also referred to herein as a "screening assay") for identifying modulators, *i.e.*, candidate or test compounds or agents (*e.g.*, peptides, peptidomimetics, small molecules or other drugs) that bind to BLAA polypeptides or have a stimulatory or inhibitory effect on, for example, BLAA expression or BLAA activity.

In one embodiment, the invention provides assays for screening candidate or test compounds which bind to or modulate the activity of a BLAA protein or polypeptide or biologically active portion thereof. The test compounds of the present invention can be obtained using any of the numerous approaches in combinatorial library methods known in the art, including: biological libraries; spatially addressable parallel solid phase or solution phase libraries; synthetic library methods requiring deconvolution; the "one-bead one-compound" library method; and synthetic library methods using affinity chromatography selection. The biological library approach is limited to peptide libraries, while the other four approaches are applicable to peptide, non-peptide oligomer or small molecule libraries of compounds (Lam
5 (1997) *Anticancer Drug Des* 12:145).

Examples of methods for the synthesis of molecular libraries can be found in the art, for example in: DeWitt *et al.* (1993) *Proc Natl Acad Sci U.S.A.* 90:6909; Erb *et al.* (1994) *Proc Natl Acad Sci U.S.A.* 91:11422; Zuckermann *et al.* (1994) *J Med Chem* 37:2678; Cho *et al.* (1993) *Science* 261:1303; Carrell *et al.* (1994) *Angew Chem Int Ed Engl* 33:2059; Carell *et al.* (1994) *Angew Chem Int Ed Engl* 33:2061; and Gallop *et al.* (1994) *J Med Chem* 37:1233.
15

Libraries of compounds may be presented in solution (*e.g.*, Houghten (1992) *Biotechniques* 13:412-421), or on beads (Lam (1991) *Nature* 354:82-84), on chips (Fodor (1993) *Nature* 364:555-556), bacteria (Ladner U.S. Pat. No. 5,223,409), spores (Ladner USP '409), plasmids (Cull *et al.* (1992) *Proc Natl Acad Sci USA* 89:1865-1869) or on phage (Scott and Smith (1990) *Science* 249:386-390; Devlin (1990) *Science* 249:404-406; Cwirla *et al.* (1990) *Proc Natl Acad Sci U.S.A.* 87:6378-6382; Felici (1991) *J Mol Biol* 222:301-310; Ladner above.).
20

In one embodiment, an assay is a cell-based assay in which a cell which expresses a form of BLAA polypeptide, or a biologically active portion thereof, on the cell surface is contacted with a test compound and the ability of the test compound to bind to a BLAA protein determined. The cell, for example, can be of mammalian origin or be a yeast cell. Determining the ability of the test compound to bind to the BLAA polypeptide can be accomplished, for example, by coupling the test compound with a radioisotope or enzymatic label such that binding of the test compound to the BLAA polypeptide or biologically active
25 portion thereof can be determined by detecting the labeled compound in a complex. For
30

example, test compounds can be labeled with ^{125}I , ^{35}S , ^{14}C , or ^3H , either directly or indirectly, and the radioisotope detected by direct counting of radioemission or by scintillation counting. Alternatively, test compounds can be enzymatically labeled with, for example, horseradish peroxidase, alkaline phosphatase, or luciferase, and the enzymatic label detected by

5 determination of conversion of an appropriate substrate to product. In one embodiment, the assay comprises contacting a cell which expresses a membrane-bound form of BLAA polypeptide, or a biologically active portion thereof, on the cell surface with a known compound which binds BLAA to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with a BLAA

10 polypeptide, wherein determining the ability of the test compound to interact with a BLAA polypeptide comprises determining the ability of the test compound to preferentially bind to BLAA or a biologically active portion thereof as compared to the known compound.

In another embodiment, an assay is a cell-based assay comprising contacting a cell expressing a membrane-bound form of BLAA polypeptide, or a biologically active portion

15 thereof, on the cell surface with a test compound and determining the ability of the test compound to modulate (*e.g.*, stimulate or inhibit) the activity of the BLAA polypeptide or biologically active portion thereof. Determining the ability of the test compound to modulate the activity of BLAA or a biologically active portion thereof can be accomplished, for example, by determining the ability of the BLAA polypeptide to bind to or interact with a

20 BLAA target molecule. As used herein, a "target molecule" is a molecule with which a BLAA polypeptide binds or interacts in nature, for example, a molecule on the surface of a cell which expresses a BLAA-interacting protein, a molecule on the surface of a second cell, a molecule in the extracellular milieu, a molecule associated with the internal surface of a cell membrane or a cytoplasmic molecule. A BLAA target molecule can be a non-BLAA molecule or a

25 BLAA protein or polypeptide of the present invention. In one embodiment, a BLAA target molecule is a component of a signal transduction pathway that facilitates transduction of an extracellular signal (*e.g.* a signal generated by binding of a compound to a membrane-bound BLAA molecule) through the cell membrane and into the cell. The target, for example, can be a second intercellular protein that has catalytic activity or a protein that facilitates the

30 association of downstream signaling molecules with BLAA.

Determining the ability of the BLAA polypeptide to bind to or interact with a BLAA target molecule can be accomplished by one of the methods described above for determining direct binding. In one embodiment, determining the ability of the BLAA polypeptide to bind to or interact with a BLAA target molecule can be accomplished by determining the activity of the target molecule. For example, the activity of the target molecule can be determined by detecting induction of a cellular second messenger of the target (*i.e.* intracellular Ca^{2+} , diacylglycerol, IP_3 , etc.), detecting catalytic/enzymatic activity of the target an appropriate substrate, detecting the induction of a reporter gene (comprising a BLAA-responsive regulatory element operatively linked to a nucleic acid encoding a detectable marker, *e.g.*, luciferase), or detecting a cellular response, for example, cell survival, cellular differentiation, or cell proliferation.

In yet another embodiment, an assay of the present invention is a cell-free assay comprising contacting a BLAA polypeptide or biologically active portion thereof with a test compound and determining the ability of the test compound to bind to the BLAA polypeptide or biologically active portion thereof. Binding of the test compound to the BLAA polypeptide can be determined either directly or indirectly as described above. In one embodiment, the assay comprises contacting the BLAA polypeptide or biologically active portion thereof with a known compound which binds BLAA to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with a BLAA polypeptide, wherein determining the ability of the test compound to interact with a BLAA polypeptide comprises determining the ability of the test compound to preferentially bind to BLAA or biologically active portion thereof as compared to the known compound.

In another embodiment, an assay is a cell-free assay comprising contacting BLAA polypeptide or biologically active portion thereof with a test compound and determining the ability of the test compound to modulate (*e.g.* stimulate or inhibit) the activity of the BLAA polypeptide or biologically active portion thereof. Determining the ability of the test compound to modulate the activity of BLAA can be accomplished, for example, by determining the ability of the BLAA polypeptide to bind to a BLAA target molecule by one of the methods described above for determining direct binding. In an alternative embodiment, determining the ability of the test compound to modulate the activity of BLAA can be

accomplished by determining the ability of the BLAA polypeptide further modulate a BLAA target molecule. For example, the catalytic/enzymatic activity of the target molecule on an appropriate substrate can be determined as previously described.

In yet another embodiment, the cell-free assay comprises contacting the BLAA
5 polypeptide or biologically active portion thereof with a known compound which binds BLAA to form an assay mixture, contacting the assay mixture with a test compound, and determining the ability of the test compound to interact with a BLAA polypeptide, wherein determining the ability of the test compound to interact with a BLAA polypeptide comprises determining the ability of the BLAA polypeptide to preferentially bind to or modulate the activity of a BLAA
10 target molecule.

The cell-free assays of the present invention are amenable to use of both the soluble form or the membrane-bound form of BLAA. In the case of cell-free assays comprising the membrane-bound form of BLAA, it may be desirable to utilize a solubilizing agent such that the membrane-bound form of BLAA is maintained in solution. Examples of such solubilizing
15 agents include non-ionic detergents such as n-octylglucoside, n-dodecylglucoside, n-dodecylmaltoside, octanoyl-N-methylglucamide, decanoyl-N-methylglucamide, Triton® X-100, Triton® X-114, Thesit®, Isotridecypoly(ethylene glycol ether)_n, N-dodecyl--N,N-dimethyl-3-ammonio-1-propane sulfonate, 3-(3-cholamidopropyl)dimethylamminiol-1-propane sulfonate (CHAPS), or 3-(3-cholamidopropyl)dimethylamminiol-2-hydroxy-
20 1-propane sulfonate (CHAPSO).

In more than one embodiment of the above assay methods of the present invention, it may be desirable to immobilize either BLAA or its target molecule to facilitate separation of complexed from uncomplexed forms of one or both of the proteins, as well as to accommodate automation of the assay. Binding of a test compound to BLAA, or interaction of BLAA with a
25 target molecule in the presence and absence of a candidate compound, can be accomplished in any vessel suitable for containing the reactants. Examples of such vessels include microtiter plates, test tubes, and micro-centrifuge tubes. In one embodiment, a fusion protein can be provided that adds a domain that allows one or both of the proteins to be bound to a matrix. For example, GST-BLAA fusion proteins or GST-target fusion proteins can be adsorbed onto
30 glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized

microtiter plates, that are then combined with the test compound or the test compound and either the non-adsorbed target protein or BLAA polypeptide, and the mixture is incubated under conditions conducive to complex formation (*e.g.*, at physiological conditions for salt and pH). Following incubation, the beads or microtiter plate wells are washed to remove any unbound components, the matrix immobilized in the case of beads, complex determined either directly or indirectly, for example, as described above. Alternatively, the complexes can be dissociated from the matrix, and the level of BLAA binding or activity determined using standard techniques.

Other techniques for immobilizing proteins on matrices can also be used in the screening assays of the invention. For example, either BLAA or its target molecule can be immobilized utilizing conjugation of biotin and streptavidin. Biotinylated BLAA or target molecules can be prepared from biotin-NHS (N-hydroxy-succinimide) using techniques well known in the art (*e.g.*, biotinylation kit, Pierce Chemicals, Rockford, Ill.), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies reactive with BLAA or target molecules, but which do not interfere with binding of the BLAA protein to its target molecule, can be derivatized to the wells of the plate, and unbound target or BLAA trapped in the wells by antibody conjugation. Methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies reactive with the BLAA or target molecule, as well as enzyme-linked assays that rely on detecting an enzymatic activity associated with the BLAA or target molecule.

In another embodiment, modulators of BLAA expression are identified in a method wherein a cell is contacted with a candidate compound and the expression of BLAA mRNA or polypeptide in the cell is determined. The level of expression of BLAA mRNA or polypeptide in the presence of the candidate compound is compared to the level of expression of BLAA mRNA or polypeptide in the absence of the candidate compound. The candidate compound can then be identified as a modulator of BLAA expression based on this comparison. For example, when expression of BLAA mRNA or polypeptide is greater (statistically significantly greater) in the presence of the candidate compound than in its absence, the candidate compound is identified as a stimulator of BLAA mRNA or polypeptide expression.

Alternatively, when expression of BLAA mRNA or polypeptide is less (statistically significantly less) in the presence of the candidate compound than in its absence, the candidate compound is identified as an inhibitor of BLAA mRNA or polypeptide expression. The level of BLAA mRNA or protein expression in the cells can be determined by methods described
5 herein for detecting BLAA mRNA or protein.

In yet another aspect of the invention, a BLAA polypeptide can be used as "bait proteins" in a two-hybrid assay or three hybrid assay (see, *e.g.*, U.S. Pat. No. 5,283,317; Zervos *et al.* (1993) *Cell* 72:223-232; Madura *et al.* (1993) *J Biol Chem* 268:12046-12054; Bartel *et al.* (1993) *Biotechniques* 14:920-924; Iwabuchi *et al.* (1993) *Oncogene* 8:1693-1696;
10 and Brent WO94/10300), to identify other proteins that bind to or interact with BLAA ("BLAA-binding proteins" or "BLAA-bp") and modulate BLAA activity. Such BLAA-binding proteins are also likely to be involved in the propagation of signals by the BLAA polypeptides as, for example, upstream or downstream elements of the BLAA pathway.

The two-hybrid system is based on the modular nature of most transcription factors,
15 which consist of separable DNA-binding and activation domains. Briefly, the assay utilizes two different DNA constructs. In one construct, the gene that codes for BLAA is fused to a gene encoding the DNA binding domain of a known transcription factor (*e.g.*, GAL-4). In the other construct, a DNA sequence, from a library of DNA sequences, that encodes an unidentified protein ("prey" or "sample") is fused to a gene that codes for the activation
20 domain of the known transcription factor. If the "bait" and the "prey" proteins are able to interact, *in vivo*, forming a BLAA-dependent complex, the DNA-binding and activation domains of the transcription factor are brought into close proximity. This proximity allows transcription of a reporter gene (*e.g.*, LacZ) that is operably linked to a transcriptional regulatory site responsive to the transcription factor. Expression of the reporter gene can be
25 detected and cell colonies containing the functional transcription factor can be isolated and used to obtain the cloned gene that encodes the protein which interacts with BLAA.

In another embodiment, modulators of activity or latency of (*i.e.*, the lack of activity) or predisposition to an immune response-associated disorder are identified wherein a test animal at increased risk for an immune response-associated disorder is administered with a test
30 compound. The test animal of this embodiment recombinantly expresses a BLAA

polypeptide. The activity of said polypeptide is then measured in the test animal. Next, the activity of the polypeptide is measured in a control animal that recombinantly expresses the polypeptide but that is not at an increased risk for an immune response-associated disorder. Finally, expression in the test animal is compared to the expression in the control animal. A
5 difference in the test animal relative to the control animal indicates that the test compound is a modulation of activity or latency of or predisposition to an immune response-associated disorder. For example, an increase in expression in the test animal indicates that the test compound is a stimulator of an immune response-associated disorder. Likewise, a decrease in the test animal indicates that the test compound is an inhibitor of an immune response-
10 associated disorder.

The test animal may be a recombinant test animal that expresses a test protein transgene, as described above, or expresses said transgene under the control of a promoter at increased levels compared to a wild type test animal. In this embodiment, the promoter is not the native promoter of the transgene.

15 This invention further pertains to novel agents identified by the above-described screening assays and uses thereof for treatments as described herein.

Detection Assays

Portions or fragments of the cDNA sequences identified herein (and the corresponding complete gene sequences) can be used in numerous ways as polynucleotide reagents. For
20 example, these sequences can be used to: (i) map their respective genes on a chromosome; and, thus, locate gene regions associated with genetic disease; and (ii) identify an individual from a minute biological sample (tissue typing). These applications are described in the subsections below.

Chromosome Mapping

25 Once the sequence (or a portion of the sequence) of a gene has been isolated, this sequence can be used to map the location of the gene on a chromosome. This process is called chromosome mapping. Accordingly, portions or fragments of the BLAA sequences described herein, can be used to map the location of the BLAA genes, respectively, on a chromosome.

The mapping of the BLAA sequences to chromosomes is an important first step in correlating these sequences with genes associated with disease.

Briefly, BLAA genes can be mapped to chromosomes by preparing PCR primers (preferably 15-25 bp in length) from the BLAA sequences. Computer analysis of the BLAA, sequences can be used to rapidly select primers that do not span more than one exon in the genomic DNA, thus complicating the amplification process. These primers can then be used for PCR screening of somatic cell hybrids containing individual human chromosomes. Only those hybrids containing the human gene corresponding to the BLAA sequences will yield an amplified fragment.

Somatic cell hybrids are prepared by fusing somatic cells from different mammals (*e.g.*, human and mouse cells). As hybrids of human and mouse cells grow and divide, they gradually lose human chromosomes in random order, but retain the mouse chromosomes. By using media in which mouse cells cannot grow, because they lack a particular enzyme, but in which human cells can, the one human chromosome that contains the gene encoding the needed enzyme will be retained. By using various media, panels of hybrid cell lines can be established. Each cell line in a panel contains either a single human chromosome or a small number of human chromosomes, and a full set of mouse chromosomes, allowing easy mapping of individual genes to specific human chromosomes. (D'Eustachio *et al.* (1983) *Science* 220:919-924). Somatic cell hybrids containing only fragments of human chromosomes can also be produced by using human chromosomes with translocations and deletions.

PCR mapping of somatic cell hybrids is a rapid procedure for assigning a particular sequence to a particular chromosome. Three or more sequences can be assigned per day using a single thermal cycler. Using the BLAA sequences to design oligonucleotide primers, sublocalization can be achieved with panels of fragments from specific chromosomes.

Fluorescence *in situ* hybridization (FISH) of a DNA sequence to a metaphase chromosomal spread can further be used to provide a precise chromosomal location in one step. Chromosome spreads can be made using cells whose division has been blocked in metaphase by a chemical like colcemid that disrupts the mitotic spindle. The chromosomes can be treated briefly with trypsin, and then stained with Giemsa. A pattern of light and dark

bands develops on each chromosome, so that the chromosomes can be identified individually. The FISH technique can be used with a DNA sequence as short as 500 or 600 bases. However, clones larger than 1,000 bases have a higher likelihood of binding to a unique chromosomal location with sufficient signal intensity for simple detection. Preferably 1,000
5 bases, and more preferably 2,000 bases, will suffice to get good results at a reasonable amount of time. For a review of this technique, see Verma *et al.*, HUMAN CHROMOSOMES: A MANUAL OF BASIC TECHNIQUES (Pergamon Press, New York 1988).

Reagents for chromosome mapping can be used individually to mark a single chromosome or a single site on that chromosome, or panels of reagents can be used for
10 marking multiple sites and/or multiple chromosomes. Reagents corresponding to noncoding regions of the genes actually are preferred for mapping purposes. Coding sequences are more likely to be conserved within gene families, thus increasing the chance of cross hybridizations during chromosomal mapping.

Once a sequence has been mapped to a precise chromosomal location, the physical
15 position of the sequence on the chromosome can be correlated with genetic map data. Such data are found, for example, in McKusick, MENDELIAN INHERITANCE IN MAN, available on-line through Johns Hopkins University Welch Medical Library). The relationship between genes and disease, mapped to the same chromosomal region, can then be identified through linkage analysis (co-inheritance of physically adjacent genes), described in, for example,
20 Egeland *et al.* (1987) *Nature*, 325:783-787.

Moreover, differences in the DNA sequences between individuals affected and unaffected with a disease associated with the BLAA gene, can be determined. If a mutation is observed in some or all of the affected individuals but not in any unaffected individuals, then the mutation is likely to be the causative agent of the particular disease. Comparison of
25 affected and unaffected individuals generally involves first looking for structural alterations in the chromosomes, such as deletions or translocations that are visible from chromosome spreads or detectable using PCR based on that DNA sequence. Ultimately, complete sequencing of genes from several individuals can be performed to confirm the presence of a mutation and to distinguish mutations from polymorphisms.

Tissue Typing

The BLAA sequences of the present invention can also be used to identify individuals from minute biological samples. In this technique, an individual's genomic DNA is digested with one or more restriction enzymes, and probed on a Southern blot to yield unique bands for
5 identification. The sequences of the present invention are useful as additional DNA markers for RFLP ("restriction fragment length polymorphisms," described in U.S. Pat. No. 5,272,057).

Furthermore, the sequences of the present invention can be used to provide an alternative technique that determines the actual base-by-base DNA sequence of selected
10 portions of an individual's genome. Thus, the BLAA sequences described herein can be used to prepare two PCR primers from the 5' and 3' ends of the sequences. These primers can then be used to amplify an individual's DNA and subsequently sequence it.

Panels of corresponding DNA sequences from individuals, prepared in this manner, can provide unique individual identifications, as each individual will have a unique set of such
15 DNA sequences due to allelic differences. The sequences of the present invention can be used to obtain such identification sequences from individuals and from tissue. The BLAA sequences of the invention uniquely represent portions of the human genome. Allelic variation occurs to some degree in the coding regions of these sequences, and to a greater degree in the noncoding regions. It is estimated that allelic variation between individual humans occurs
20 with a frequency of about once per each 500 bases. Much of the allelic variation is due to single nucleotide polymorphisms (SNPs), which include restriction fragment length polymorphisms (RFLPs).

Each of the sequences described herein can, to some degree, be used as a standard against which DNA from an individual can be compared for identification purposes. Because
25 greater numbers of polymorphisms occur in the noncoding regions, fewer sequences are necessary to differentiate individuals. The noncoding sequences of SEQ ID NO: 1, SEQ ID NO: 3, and SEQ ID NO: 5 (SEQ ID NOs: 8, 10, and 12, respectively) can comfortably provide positive individual identification with a panel of perhaps 10 to 1,000 primers that each yield a noncoding amplified sequence of 100 bases. If predicted coding sequences, such as those in

SEQ ID NO: 1, 3, 5, 7, 9, and 11 are used, a more appropriate number of primers for individual positive identification would be 500-2000.

Predictive Medicine

The present invention also pertains to the field of predictive medicine in which
5 diagnostic assays, prognostic assays, pharmacogenomics, and monitoring clinical trials are used for prognostic (predictive) purposes to thereby treat an individual prophylactically. Accordingly, one aspect of the present invention relates to diagnostic assays for determining BLAA polypeptide and/or nucleic acid expression as well as BLAA activity, in the context of a biological sample (*e.g.*, blood, serum, cells, tissue) to thereby determine whether an
10 individual is afflicted with a disease or disorder, or is at risk of developing a disorder, associated with aberrant BLAA expression or activity. The invention also provides for prognostic (or predictive) assays for determining whether an individual is at risk of developing a disorder associated with BLAA polypeptide or nucleic acid expression or activity. For example, mutations in a BLAA gene can be assayed in a biological sample. Such assays can
15 be used for prognostic or predictive purpose to thereby prophylactically treat an individual prior to the onset of a disorder characterized by or associated with BLAA polypeptide or nucleic acid expression or activity.

Another aspect of the invention provides methods for determining BLAA polypeptide or nucleic acid expression or BLAA activity in an individual to thereby select appropriate
20 therapeutic or prophylactic agents for that individual (referred to herein as "pharmacogenomics"). Pharmacogenomics allows for the selection of agents (*e.g.*, drugs) for therapeutic or prophylactic treatment of an individual based on the genotype of the individual (*e.g.*, the genotype of the individual examined to determine the ability of the individual to respond to a particular agent.)

25 Yet another aspect of the invention pertains to monitoring the influence of agents (*e.g.*, drugs, compounds) on the expression or activity of BLAA in clinical trials.

These and other agents are described in further detail in the following sections.

Diagnostic Assays

An exemplary method for detecting the presence or absence of BLAA in a biological sample involves obtaining a biological sample from a test subject and contacting the biological sample with a compound or an agent capable of detecting BLAA polypeptide or nucleic acid (e.g., mRNA, genomic DNA) that encodes BLAA polypeptide such that the presence of BLAA is detected in the biological sample. An agent for detecting BLAA mRNA or genomic DNA is a labeled nucleic acid probe capable of hybridizing to BLAA mRNA or genomic DNA. The nucleic acid probe can be, for example, a full-length BLAA nucleic acid, such as the nucleic acid of SEQ ID NO: 1, 3, 5, 7, 8, 9, 10, 11, or 12, or a portion thereof, such as an oligonucleotide of at least 15, 30, 50, 100, 250 or 500 nucleotides in length and sufficient to specifically hybridize under stringent conditions to BLAA mRNA or genomic DNA. Other suitable probes for use in the diagnostic assays of the invention are described herein.

An agent for detecting BLAA polypeptide is an antibody capable of binding to BLAA protein, preferably an antibody with a detectable label. Antibodies can be polyclonal, or more preferably, monoclonal. An intact antibody, or a fragment thereof (e.g., Fab or F(ab')₂) can be used. The term "labeled", with regard to the probe or antibody, is intended to encompass direct labeling of the probe or antibody by coupling (i.e., physically linking) a detectable substance to the probe or antibody, as well as indirect labeling of the probe or antibody by reactivity with another reagent that is directly labeled. Examples of indirect labeling include detection of a primary antibody using a fluorescently labeled secondary antibody and end-labeling of a DNA probe with biotin such that it can be detected with fluorescently labeled streptavidin. The term "biological sample" is intended to include tissues, cells and biological fluids isolated from a subject, as well as tissues, cells and fluids present within a subject. That is, the detection method of the invention can be used to detect BLAA mRNA, polypeptide, or genomic DNA in a biological sample *in vitro* as well as *in vivo*. For example, *in vitro* techniques for detection of BLAA mRNA include Northern hybridizations and *in situ* hybridizations. *In vitro* techniques for detection of BLAA polypeptide include enzyme linked immunosorbent assays (ELISAs), Western blots, immunoprecipitations and immunofluorescence. *In vitro* techniques for detection of BLAA genomic DNA include Southern hybridizations. Furthermore, *in vivo* techniques for detection of BLAA protein

include introducing into a subject a labeled anti-BLAA antibody. For example, the antibody can be labeled with a radioactive marker whose presence and location in a subject can be detected by standard imaging techniques.

In one embodiment, the biological sample contains protein molecules from the test subject. Alternatively, the biological sample can contain mRNA molecules from the test subject or genomic DNA molecules from the test subject. A preferred biological sample is a peripheral blood leukocyte sample isolated by conventional means from a subject.

In another embodiment, the methods further involve obtaining a control biological sample from a control subject, contacting the control sample with a compound or agent capable of detecting BLAA polypeptide, mRNA, or genomic DNA, such that the presence of BLAA polypeptide, mRNA or genomic DNA is detected in the biological sample, and comparing the presence of BLAA polypeptide, mRNA or genomic DNA in the control sample with the presence of BLAA polypeptide, mRNA or genomic DNA in the test sample.

The invention also encompasses kits for detecting the presence of BLAA in a biological sample. For example, the kit can comprise: a labeled compound or agent capable of detecting BLAA polypeptide or mRNA in a biological sample; means for determining the amount of BLAA in the sample; and means for comparing the amount of BLAA in the sample with a standard. The compound or agent can be packaged in a suitable container. The kit can further comprise instructions for using the kit to detect BLAA polypeptide or nucleic acid.

Prognostic Assays

The diagnostic methods described herein can furthermore be utilized to identify subjects having or at risk of developing a disease or disorder associated with aberrant BLAA expression or activity. For example, the assays described herein, such as the preceding diagnostic assays or the following assays, can be utilized to identify a subject having or at risk of developing an immune response-associated disorder associated with BLAA polypeptide or nucleic acid expression or activity such as cancers, infectious diseases, autoimmune disorders, and complications associated with transplantation. Alternatively, the prognostic assays can be utilized to identify a subject having or at risk for developing a disease or disorder. Thus, the present invention provides a method for identifying a disease or disorder associated with aberrant BLAA expression or activity in which a test sample is obtained from a subject and

BLAA polypeptide or nucleic acid (*e.g.*, mRNA, genomic DNA) is detected, wherein the presence of BLAA polypeptide or nucleic acid is diagnostic for a subject having or at risk of developing a disease or disorder associated with aberrant BLAA expression or activity. As used herein, a "test sample" refers to a biological sample obtained from a subject of interest.

5 For example, a test sample can be a biological fluid (*e.g.*, serum), cell sample, or tissue.

Furthermore, the prognostic assays described herein can be used to determine whether a subject can be administered an agent (*e.g.*, an agonist, antagonist, peptidomimetic, protein, peptide, nucleic acid, small molecule, or other drug candidate) to treat a disease or disorder associated with aberrant BLAA expression or activity. For example, such methods can be
10 used to determine whether a subject can be effectively treated with an agent for a disorder, such as cancer, infectious disease, autoimmune disorders, and complications associated with transplantation. Thus, the present invention provides methods for determining whether a subject can be effectively treated with an agent for a disorder associated with aberrant BLAA expression or activity in which a test sample is obtained and a BLAA polypeptide or nucleic
15 acid is detected (*e.g.*, wherein the presence of a BLAA polypeptide or nucleic acid is diagnostic for a subject that can be administered the agent to treat a disorder associated with aberrant BLAA expression or activity.)

The methods of the invention can also be used to detect genetic lesions in a BLAA gene, thereby determining if a subject with the lesioned gene is at risk for a disorder
20 characterized by aberrant cell proliferation and/or differentiation. In various embodiments, the methods include detecting, in a sample of cells from the subject, the presence or absence of a genetic lesion characterized by at least one of an alteration affecting the integrity of a gene encoding a BLAA polypeptide, or the mis-expression of the BLAA gene. For example, such genetic lesions can be detected by ascertaining the existence of at least one of (1) a deletion of
25 one or more nucleotides from a BLAA gene; (2) an addition of one or more nucleotides to a BLAA gene; (3) a substitution of one or more nucleotides of a BLAA gene, (4) a chromosomal rearrangement of a BLAA gene; (5) an alteration in the level of a messenger RNA transcript of a BLAA gene, (6) aberrant modification of a BLAA gene, such as of the methylation pattern of the genomic DNA, (7) the presence of a non-wild type splicing pattern of a messenger RNA
30 transcript of a BLAA gene, (8) a non-wild type level of a BLAA polypeptide, (9) allelic loss of

a BLAA gene, and (10) inappropriate post-translational modification of a BLAA polypeptide. As described herein, there are a large number of assay techniques known in the art which can be used for detecting lesions in a BLAA gene. A preferred biological sample is a peripheral blood leukocyte sample isolated by conventional means from a subject. However, any
5 biological sample containing nucleated cells may be used, including, for example, buccal mucosal cells.

In certain embodiments, detection of the lesion involves the use of a probe/primer in a polymerase chain reaction (PCR) (see, *e.g.*, U.S. Pat. Nos. 4,683,195 and 4,683,202), such as anchor PCR or RACE PCR, or, alternatively, in a ligation chain reaction (LCR) (see, *e.g.*,
10 Landegran *et al.* (1988) *Science* 241:1077-1080; and Nakazawa *et al.* (1994) *PNAS* 91:360-364), the latter of which can be particularly useful for detecting point mutations in the BLAA-gene (see Abravaya *et al.* (1995) *Nucl Acids Res* 23:675-682). This method can include the steps of collecting a sample of cells from a patient, isolating nucleic acid (*e.g.*, genomic, mRNA or both) from the cells of the sample, contacting the nucleic acid sample with one or
15 more primers that specifically hybridize to a BLAA gene under conditions such that hybridization and amplification of the BLAA gene (if present) occurs, and detecting the presence or absence of an amplification product, or detecting the size of the amplification product and comparing the length to a control sample. It is anticipated that PCR and/or LCR may be desirable to use as a preliminary amplification step in conjunction with any of the
20 techniques used for detecting mutations described herein.

Alternative amplification methods include: self sustained sequence replication (Guatelli *et al.*, 1990, *Proc Natl Acad Sci USA* 87:1874-1878), transcriptional amplification system (Kwoh, *et al.*, 1989, *Proc Natl Acad Sci USA* 86:1173-1177), Q-Beta Replicase (Lizardi *et al.*, 1988, *BioTechnology* 6:1197), or any other nucleic acid amplification method,
25 followed by the detection of the amplified molecules using techniques well known to those of skill in the art. These detection schemes are especially useful for the detection of nucleic acid molecules if such molecules are present in very low numbers.

In an alternative embodiment, mutations in a BLAA gene from a sample cell can be identified by alterations in restriction enzyme cleavage patterns. For example, sample and
30 control DNA is isolated, amplified (optionally), digested with one or more restriction

endonucleases, and fragment length sizes are determined by gel electrophoresis and compared. Differences in fragment length sizes between sample and control DNA indicates mutations in the sample DNA. Moreover, the use of sequence specific ribozymes (see, for example, U.S. Pat. No. 5,493,531) can be used to score for the presence of specific mutations by development
5 or loss of a ribozyme cleavage site.

In other embodiments, genetic mutations in BLAA can be identified by hybridizing a sample and control nucleic acids, *e.g.*, DNA or RNA, to high density arrays containing hundreds or thousands of oligonucleotides probes (Cronin *et al.* (1996) *Human Mutation* 7: 244-255; Kozal *et al.* (1996) *Nature Medicine* 2: 753-759). For example, genetic mutations in
10 BLAA can be identified in two dimensional arrays containing light-generated DNA probes as described in Cronin *et al.* above. Briefly, a first hybridization array of probes can be used to scan through long stretches of DNA in a sample and control to identify base changes between the sequences by making linear arrays of sequential overlapping probes. This step allows the identification of point mutations. This step is followed by a second hybridization array that
15 allows the characterization of specific mutations by using smaller, specialized probe arrays complementary to all variants or mutations detected. Each mutation array is composed of parallel probe sets, one complementary to the wild-type gene and the other complementary to the mutant gene.

In yet another embodiment, any of a variety of sequencing reactions known in the art
20 can be used to directly sequence the BLAA gene and detect mutations by comparing the sequence of the sample BLAA with the corresponding wild-type (control) sequence. Examples of sequencing reactions include those based on techniques developed by Maxim and Gilbert (1977) *PNAS* 74:560 or Sanger (1977) *PNAS* 74:5463. It is also contemplated that any of a variety of automated sequencing procedures can be utilized when performing the
25 diagnostic assays (Naeve *et al.*, (1995) *Biotechniques* 19:448), including sequencing by mass spectrometry (see, *e.g.*, PCT International Publ. No. WO 94/16101; Cohen *et al.* (1996) *Adv Chromatogr* 36:127-162; and Griffin *et al.* (1993) *Appl Biochem Biotechnol* 38:147-159).

Other methods for detecting mutations in the BLAA gene include methods in which protection from cleavage agents is used to detect mismatched bases in RNA/RNA or
30 RNA/DNA heteroduplexes (Myers *et al.* (1985) *Science* 230:1242). In general, the art

technique of "mismatch cleavage" starts by providing heteroduplexes of formed by hybridizing (labeled) RNA or DNA containing the wild-type BLAA sequence with potentially mutant RNA or DNA obtained from a tissue sample. The double-stranded duplexes are treated with an agent that cleaves single-stranded regions of the duplex such as which will exist due to basepair mismatches between the control and sample strands. For instance, RNA/DNA duplexes can be treated with RNase and DNA/DNA hybrids treated with S1 nuclease to enzymatically digesting the mismatched regions. In other embodiments, either DNA/DNA or RNA/DNA duplexes can be treated with hydroxylamine or osmium tetroxide and with piperidine in order to digest mismatched regions. After digestion of the mismatched regions, the resulting material is then separated by size on denaturing polyacrylamide gels to determine the site of mutation. See, for example, Cotton *et al* (1988) *Proc Natl Acad Sci USA* 85:4397; Saleeba *et al* (1992) *Methods Enzymol* 217:286-295. In an embodiment, the control DNA or RNA can be labeled for detection.

In still another embodiment, the mismatch cleavage reaction employs one or more proteins that recognize mismatched base pairs in double-stranded DNA (so called "DNA mismatch repair" enzymes) in defined systems for detecting and mapping point mutations in BLAA cDNAs obtained from samples of cells. For example, the mutY enzyme of *E. coli* cleaves A at G/A mismatches and the thymidine DNA glycosylase from HeLa cells cleaves T at G/T mismatches (Hsu *et al.* (1994) *Carcinogenesis* 15:1657-1662). According to an exemplary embodiment, a probe based on a BLAA sequence, *e.g.*, a wild-type BLAA sequence, is hybridized to a cDNA or other DNA product from a test cell(s). The duplex is treated with a DNA mismatch repair enzyme, and the cleavage products, if any, can be detected from electrophoresis protocols or the like. See, for example, U.S. Pat. No. 5,459,039.

In other embodiments, alterations in electrophoretic mobility will be used to identify mutations in BLAA genes. For example, single strand conformation polymorphism (SSCP) may be used to detect differences in electrophoretic mobility between mutant and wild type nucleic acids (Orita *et al.* (1989) *Proc Natl Acad Sci USA*: 86:2766, see also Cotton (1993) *Mutat Res* 285:125-144; Hayashi (1992) *Genet Anal Tech Appl* 9:73-79). Single-stranded DNA fragments of sample and control BLAA nucleic acids will be denatured and allowed to renature. The secondary structure of single-stranded nucleic acids varies according to

sequence, the resulting alteration in electrophoretic mobility enables the detection of even a single base change. The DNA fragments may be labeled or detected with labeled probes. The sensitivity of the assay may be enhanced by using RNA (rather than DNA), in which the secondary structure is more sensitive to a change in sequence. In one embodiment, the subject
5 method utilizes heteroduplex analysis to separate double stranded heteroduplex molecules on the basis of changes in electrophoretic mobility (Keen *et al.* (1991) *Trends Genet* 7:5).

In yet another embodiment the movement of mutant or wild-type fragments in polyacrylamide gels containing a gradient of denaturant is assayed using denaturing gradient gel electrophoresis (DGGE) (Myers *et al.* (1985) *Nature* 313:495). When DGGE is used as the
10 method of analysis, DNA will be modified to insure that it does not completely denature, for example by adding a GC clamp of approximately 40 bp of high-melting GC-rich DNA by PCR. In a further embodiment, a temperature gradient is used in place of a denaturing gradient to identify differences in the mobility of control and sample DNA (Rosenbaum and Reissner (1987) *Biophys Chem* 265:12753).

15 Examples of other techniques for detecting point mutations include, but are not limited to, selective oligonucleotide hybridization, selective amplification, or selective primer extension. For example, oligonucleotide primers may be prepared in which the known mutation is placed centrally and then hybridized to target DNA under conditions that permit hybridization only if a perfect match is found (Saiki *et al.* (1986) *Nature* 324:163); Saiki *et al.*
20 (1989) *Proc Natl Acad. Sci USA* 86:6230). Such allele specific oligonucleotides are hybridized to PCR amplified target DNA or a number of different mutations when the oligonucleotides are attached to the hybridizing membrane and hybridized with labeled target DNA.

Alternatively, allele specific amplification technology that depends on selective PCR
25 amplification may be used in conjunction with the instant invention. Oligonucleotides used as primers for specific amplification may carry the mutation of interest in the center of the molecule (so that amplification depends on differential hybridization) (Gibbs *et al.* (1989) *Nucleic Acids Res* 17:2437-2448) or at the extreme 3' end of one primer where, under appropriate conditions, mismatch can prevent, or reduce polymerase extension (Prossner
30 (1993) *Tibtech* 11:238). In addition it may be desirable to introduce a novel restriction site in

the region of the mutation to create cleavage-based detection (Gasparini *et al* (1992) *Mol Cell Probes* 6:1). It is anticipated that in certain embodiments amplification may also be performed using Taq ligase for amplification (Barany (1991) *Proc Natl Acad Sci USA* 88:189). In such cases, ligation will occur only if there is a perfect match at the 3' end of the 5' sequence, making it possible to detect the presence of a known mutation at a specific site by looking for the presence or absence of amplification.

The methods described herein may be performed, for example, by utilizing pre-packaged diagnostic kits comprising at least one probe nucleic acid or antibody reagent described herein, which may be conveniently used, *e.g.*, in clinical settings to diagnose patients exhibiting symptoms or family history of a disease or illness involving a BLAA gene.

Furthermore, any cell type or tissue, preferably peripheral blood leukocytes, in which BLAA is expressed may be utilized in the prognostic assays described herein. However, any biological sample containing nucleated cells may be used, including, for example, buccal mucosal cells.

15 **Pharmacogenomics**

Agents, or modulators that have a stimulatory or inhibitory effect on BLAA activity (*e.g.*, BLAA gene expression), as identified by a screening assay described herein can be administered to individuals to treat (prophylactically or therapeutically) disorders (*e.g.*, cancer, infectious disease, autoimmune disorders, and complications associated with transplantation) associated with aberrant BLAA activity. In conjunction with such treatment, the pharmacogenomics (*i.e.*, the study of the relationship between an individual's genotype and that individual's response to a foreign compound or drug) of the individual may be considered. Differences in metabolism of therapeutics can lead to severe toxicity or therapeutic failure by altering the relation between dose and blood concentration of the pharmacologically active drug. Thus, the pharmacogenomics of the individual permits the selection of effective agents (*e.g.*, drugs) for prophylactic or therapeutic treatments based on a consideration of the individual's genotype. Such pharmacogenomics can further be used to determine appropriate dosages and therapeutic regimens. Accordingly, the activity of BLAA polypeptide, expression of BLAA nucleic acid, or mutation content of BLAA genes in an individual can be determined

to thereby select appropriate agent(s) for therapeutic or prophylactic treatment of the individual.

Pharmacogenomics deals with clinically significant hereditary variations in the response to drugs due to altered drug disposition and abnormal action in affected persons. See
5 *e.g.*, Eichelbaum, *Clin Exp Pharmacol Physiol*, 1996, 23:983-985 and Linder, *Clin Chem*,
1997, 43:254-266. In general, two types of pharmacogenetic conditions can be differentiated. Genetic conditions transmitted as a single factor altering the way drugs act on the body (altered drug action) or genetic conditions transmitted as single factors altering the way the body acts on drugs (altered drug metabolism). These pharmacogenetic conditions can occur
10 either as rare defects or as polymorphisms. For example, glucose-6-phosphate dehydrogenase (G6PD) deficiency is a common inherited enzymopathy in which the main clinical complication is haemolysis after ingestion of oxidant drugs (anti-malarials, sulfonamides, analgesics, nitrofurans) and consumption of fava beans.

As an illustrative embodiment, the activity of drug metabolizing enzymes is a major
15 determinant of both the intensity and duration of drug action. The discovery of genetic polymorphisms of drug metabolizing enzymes (*e.g.*, N-acetyltransferase 2 (NAT 2) and cytochrome P450 enzymes CYP2D6 and CYP2C19) has provided an explanation as to why some patients do not obtain the expected drug effects or show exaggerated drug response and serious toxicity after taking the standard and safe dose of a drug. These polymorphisms are
20 expressed in two phenotypes in the population, the extensive metabolizer (EM) and poor metabolizer (PM). The prevalence of PM is different among different populations. For example, the gene coding for CYP2D6 is highly polymorphic and several mutations have been identified in PM, which all lead to the absence of functional CYP2D6. Poor metabolizers of CYP2D6 and CYP2C19 quite frequently experience exaggerated drug response and side
25 effects when they receive standard doses. If a metabolite is the active therapeutic moiety, PM show no therapeutic response, as demonstrated for the analgesic effect of codeine mediated by its CYP2D6-formed metabolite morphine. The other extreme are the so called ultra-rapid metabolizers who do not respond to standard doses. Recently, the molecular basis of ultra-rapid metabolism has been identified to be due to CYP2D6 gene amplification.

Thus, the activity of BLAA polypeptide, expression of BLAA nucleic acid, or mutation content of BLAA genes in an individual can be determined to thereby select appropriate agent(s) for therapeutic or prophylactic treatment of the individual. In addition, pharmacogenetic studies can be used to apply genotyping of polymorphic alleles encoding drug-metabolizing enzymes to the identification of an individual's drug responsiveness phenotype. This knowledge, when applied to dosing or drug selection, can avoid adverse reactions or therapeutic failure and thus enhance therapeutic or prophylactic efficiency when treating a subject with a BLAA modulator, such as a modulator identified by one of the exemplary screening assays described herein.

10 **Monitoring of Effects During Clinical Trials**

Monitoring the influence of agents (*e.g.*, drugs, compounds) on the expression or activity of BLAA (*e.g.*, the ability to modulate aberrant cell proliferation and/or differentiation) can be applied not only in basic drug screening, but also in clinical trials. For example, the effectiveness of an agent determined by a screening assay as described herein to increase BLAA gene expression, protein levels, or upregulate BLAA activity, can be monitored in clinical trials of subjects exhibiting decreased BLAA gene expression, protein levels, or downregulated BLAA activity. Alternatively, the effectiveness of an agent determined by a screening assay to decrease BLAA gene expression, protein levels, or downregulate BLAA activity, can be monitored in clinical trials of subjects exhibiting increased BLAA gene expression, protein levels, or upregulated BLAA activity. In such clinical trials, the expression or activity of BLAA and, preferably, other genes that have been implicated in, for example, an immune response-associated disorder can be used as a "read out" or markers of the immune responsiveness of a particular cell.

For example, and not by way of limitation, genes, including BLAA, that are modulated in cells by treatment with an agent (*e.g.*, compound, drug or small molecule) that modulates BLAA activity (*e.g.*, identified in a screening assay as described herein) can be identified. Thus, to study the effect of agents on cellular proliferation disorders, for example, in a clinical trial, cells can be isolated and RNA prepared and analyzed for the levels of expression of BLAA and other genes implicated in the disorder. The levels of gene expression (*i.e.*, a gene expression pattern) can be quantified by Northern blot analysis or RT-PCR, as described

herein, or alternatively by measuring the amount of protein produced, by one of the methods as described herein, or by measuring the levels of activity of BLAA or other genes. In this way, the gene expression pattern can serve as a marker, indicative of the physiological response of the cells to the agent. Accordingly, this response state may be determined before, and at
5 various points during, treatment of the individual with the agent.

In one embodiment, the present invention provides a method for monitoring the effectiveness of treatment of a subject with an agent (*e.g.*, an agonist, antagonist, protein, polypeptide, peptidomimetic, nucleic acid, small molecule, or other drug candidate identified by the screening assays described herein) comprising the steps of (i) obtaining a
10 pre-administration sample from a subject prior to administration of the agent; (ii) detecting the level of expression of a BLAA polypeptide, mRNA, or genomic DNA in the preadministration sample; (iii) obtaining one or more post-administration samples from the subject; (iv) detecting the level of expression or activity of the BLAA polypeptide, mRNA, or genomic DNA in the post-administration samples; (v) comparing the level of expression or activity of the BLAA
15 polypeptide, mRNA, or genomic DNA in the pre-administration sample with the BLAA polypeptide, mRNA, or genomic DNA in the post administration sample or samples; and (vi) altering the administration of the agent to the subject accordingly. For example, increased administration of the agent may be desirable to increase the expression or activity of BLAA to higher levels than detected, *i.e.*, to increase the effectiveness of the agent. Alternatively,
20 decreased administration of the agent may be desirable to decrease expression or activity of BLAA to lower levels than detected, *i.e.*, to decrease the effectiveness of the agent.

Methods of Treatment

The present invention provides for both prophylactic and therapeutic methods of treating a subject at risk of (or susceptible to) a disorder or having a disorder associated with
25 aberrant BLAA expression or activity.

Disorders

Diseases and disorders that are characterized by increased (relative to a subject not suffering from the disease or disorder) levels or biological activity may be treated with therapeutics that antagonize (*i.e.*, reduce or inhibit) activity. Therapeutics that antagonize

activity may be administered in a therapeutic or prophylactic manner. Therapeutics that may be utilized include, but are not limited to, (i) an aforementioned polypeptide, or analogs, derivatives, fragments or homologs thereof; (ii) antibodies to an aforementioned polypeptide; (iii) nucleic acids encoding an aforementioned polypeptide; (iv) administration of antisense nucleic acid and nucleic acids that are "dysfunctional" (*i.e.*, due to a heterologous insertion within the coding sequences of coding sequences to an aforementioned polypeptide) that are utilized to "knockout" endogenous function of an aforementioned peptide by homologous recombination (see, *e.g.*, Capecchi, 1989, *Science* 244: 1288-1292); or (v) modulators (*i.e.*, inhibitors, agonists and antagonists, including additional polypeptide mimetic of the invention or antibodies specific to a peptide of the invention) that alter the interaction between an aforementioned polypeptide and its binding partner.

Diseases and disorders that are characterized by decreased (relative to a subject not suffering from the disease or disorder) levels or biological activity may be treated with therapeutics that increase (*i.e.*, are agonists to) activity. Therapeutics that upregulate activity may be administered in a therapeutic or prophylactic manner. Therapeutics that may be utilized include, but are not limited to, an aforementioned polypeptide, or analogs, derivatives, fragments or homologs thereof; or an agonist that increases bioavailability.

Increased or decreased levels can be readily detected by quantifying polypeptide and/or RNA, by obtaining a patient tissue sample (*e.g.*, from biopsy tissue) and assaying it *in vitro* for RNA or peptide levels, structure and/or activity of the expressed polypeptides (or mRNAs of an aforementioned polypeptide). Methods that are well-known within the art include, but are not limited to, immunoassays (*e.g.*, by Western blot analysis, immunoprecipitation followed by sodium dodecyl sulfate (SDS) polyacrylamide gel electrophoresis, immunocytochemistry, etc.) and/or hybridization assays to detect expression of mRNAs (*e.g.*, Northern assays, dot blots, *in situ* hybridization, etc.).

Prophylactic Methods

In one aspect, the invention provides a method for preventing, in a subject, a disease or condition associated with an aberrant BLAA expression or activity, by administering to the subject an agent that modulates BLAA expression or at least one BLAA activity. Subjects at risk for a disease that is caused or contributed to by aberrant BLAA expression or activity can

be identified by, for example, any or a combination of diagnostic or prognostic assays as described herein. Administration of a prophylactic agent can occur prior to the manifestation of symptoms characteristic of the BLAA aberrancy, such that a disease or disorder is prevented or, alternatively, delayed in its progression. Depending on the type of BLAA aberrancy, for example, a BLAA agonist or BLAA antagonist agent can be used for treating the subject. The appropriate agent can be determined based on screening assays described herein. The prophylactic methods of the present invention are further discussed in the following subsections.

Therapeutic Methods

Another aspect of the invention pertains to methods of modulating BLAA expression or activity for therapeutic purposes. The modulatory method of the invention involves contacting a cell with an agent that modulates one or more of the activities of BLAA polypeptide activity associated with the cell. An agent that modulates BLAA polypeptide activity can be an agent as described herein, such as a nucleic acid or a protein, a naturally-occurring cognate ligand of a BLAA polypeptide, a peptide, a BLAA peptidomimetic, or other small molecule. In one embodiment, the agent stimulates one or more BLAA polypeptide activity. Examples of such stimulatory agents include active BLAA polypeptide and a nucleic acid molecule encoding BLAA that has been introduced into the cell. In another embodiment, the agent inhibits one or more BLAA polypeptide activity. Examples of such inhibitory agents include antisense BLAA nucleic acid molecules and anti-BLAA antibodies. These modulatory methods can be performed *in vitro* (e.g., by culturing the cell with the agent) or, alternatively, *in vivo* (e.g., by administering the agent to a subject). As such, the present invention provides methods of treating an individual afflicted with a disease or disorder characterized by aberrant expression or activity of a BLAA polypeptide or nucleic acid molecule. In one embodiment, the method involves administering an agent (e.g., an agent identified by a screening assay described herein), or combination of agents that modulates (e.g., upregulates or downregulates) BLAA expression or activity. In another embodiment, the method involves administering a BLAA polypeptide or nucleic acid molecule as therapy to compensate for reduced or aberrant BLAA expression or activity.

Stimulation of BLAA activity is desirable in situations in which BLAA is abnormally downregulated and/or in which increased BLAA activity is likely to have a beneficial effect. One example of such a situation is where a subject has a disorder characterized by aberrant cell proliferation and/or differentiation (*e.g.*, cancer). Another example of such a situation is where the subject has an immune response-associated disorder (*e.g.*, autoimmune disorders, infectious diseases, and complications associated with transplantation).

In one embodiment, this invention involves a method of treating a pathological state in a mammal comprising administering a therapeutic amount of a polypeptide that is at least 95% identical to a polypeptide with an amino acid sequence of SEQ ID NO: 2 or SEQ ID NO: 4, or a biologically active fragment thereof. An alternative embodiment involves administering to a subject an antibody that selectively binds to a BLAA polypeptide, and fragments, homologs, analogs, and derivatives thereof.

Determination of the Biological Effect of the Therapeutic

In various embodiments of the present invention, suitable *in vitro* or *in vivo* assays are utilized to determine the effect of a specific therapeutic and whether its administration is indicated for treatment of the affected tissue.

In various specific embodiments, *in vitro* assays may be performed with representative cells of the type(s) involved in the patient's disorder, to determine if a given therapeutic exerts the desired effect upon the cell type(s). Compounds for use in therapy may be tested in suitable animal model systems including, but not limited to rats, mice, chicken, cows, monkeys, rabbits, and the like, prior to testing in human subjects. Similarly, for *in vivo* testing, any of the animal model system known in the art may be used prior to administration to human subjects.

Malignancies

An aforementioned BLAA polypeptide may be involved in the regulation of cell proliferation. Accordingly, therapeutics of the present invention may be useful in the therapeutic or prophylactic treatment of diseases or disorders that are associated with cell hyperproliferation and/or loss of control of cell proliferation (*e.g.*, cancers, malignancies and

tumors). For a review of such hyperproliferation disorders, see *e.g.*, Fishman, *et al.*, 1985. MEDICINE, 2nd ed., J.B. Lippincott Co., Philadelphia, PA.

Therapeutics of the present invention may be assayed by any method known within the art for determining efficacy in treating or preventing malignancies and related disorders. Such assays include, but are not limited to, *in vitro* assays utilizing transformed cells or cells derived from the patient's tumor, as well as *in vivo* assays using animal models of cancer or malignancies. Potentially effective therapeutics are those that, for example, inhibit the proliferation of tumor-derived or transformed cells in culture or cause a regression of tumors in animal models, in comparison to the controls.

In the practice of the present invention, once a malignancy or cancer has been shown to be amenable to treatment by modulating (*i.e.*, inhibiting, antagonizing, or agonizing) activity, that cancer or malignancy may subsequently be treated or prevented by the administration of a therapeutic that serves to modulate polypeptide function.

Disorders related to organ transplantation

BLAA has been implicated in disorders related to organ transplantation, in particular, but not limited to, organ rejection. Therapeutics of the invention, particularly those that modulate (or supply) activity, may be effective in treating or preventing diseases or disorders related to organ transplantation. Therapeutics of the invention (particularly therapeutics that modulate the levels or activity of an aforementioned protein) can be assayed by any method known in the art for efficacy in treating or preventing such diseases and disorders related to organ transplantation. Such assays include *in vitro* assays for using cell culture models as described below, or *in vivo* assays using animal models of diseases and disorders related to organ transplantation, see *e.g.*, below. Potentially effective therapeutics, for example, but not by way of limitation, reduce immune rejection responses in animal models in comparison to controls.

Accordingly, once diseases and disorders related to organ transplantation are shown to be amenable to treatment by modulation of activity, such diseases or disorders can be treated or prevented by administration of a therapeutic that modulates activity.

T lymphocyte Growth and Activation Cell Proliferation/Differentiation Activity

The BLAA polypeptide disclosed in the instant invention is a new B7 family member. B7 family members, such as B7-1 and B7-2, are members of the immunoglobulin superfamily and bind to activated T lymphocytes and provide regulatory signals for T lymphocyte cell growth than activation.

Immune Stimulating or Suppressing Activity

A BLAA polypeptide of the present invention may also exhibit immune stimulating or immune suppressing activity, including without limitation the activities for which assays are described herein. A protein may be useful in the treatment of various immune deficiencies and disorders (including severe combined immunodeficiency (SCID)), *e.g.*, in regulating (up or down) growth and proliferation of T and/or B lymphocytes, as well as effecting the cytolytic activity of NK cells and other cell populations. These immune deficiencies may be genetic or be caused by viral (*e.g.*, HIV) as well as bacterial or fungal infections, or may result from autoimmune disorders. More specifically, infectious diseases caused by viral, bacterial, fungal or other infection may be treatable using a polypeptide of the present invention, including infections by HIV, hepatitis viruses, herpesviruses, mycobacteria, Leishmania species, malaria species, and various fungal infections such as candidiasis. Of course, in this regard, a polypeptide of the present invention may also be useful where a boost to the immune system generally may be desirable, *i.e.*, in the treatment of cancer.

Autoimmune disorders which may be treated using a polypeptide of the present invention include, for example, connective tissue disease, multiple sclerosis, systemic lupus erythematosus, rheumatoid arthritis, autoimmune pulmonary inflammation, Guillain-Barre syndrome, autoimmune thyroiditis, insulin dependent diabetes mellitus, myasthenia gravis, graft-versus-host disease and autoimmune inflammatory eye disease. Such a polypeptide of the present invention may also be useful in the treatment of allergic reactions and conditions, such as asthma (particularly allergic asthma) or other respiratory problems. Other conditions, in which immune suppression is desired (including, for example, organ transplantation), may also be treatable using a polypeptide of the present invention.

Using the polypeptide of the invention to modulate immune responses in a number of ways. Down regulation may be in the form of inhibiting or blocking an immune response

already in progress or may involve preventing the induction of an immune response. The functions of activated T cells may be inhibited by suppressing T cell responses or by inducing specific tolerance in T cells, or both. Immunosuppression of T cell responses is generally an active, non-antigen-specific, process which requires continuous exposure of the T cells to the suppressive agent. Tolerance, which involves inducing non-responsiveness or energy in T cells, is distinguishable from immunosuppression in that it is generally antigen-specific and persists after exposure to the tolerizing agent has ceased. Operationally, tolerance can be demonstrated by the lack of a T cell response upon re-exposure to specific antigen in the absence of the tolerizing agent.

Down regulating or preventing one or more antigen functions (including without limitation B lymphocyte antigen functions (such as, for example, B7)), *e.g.*, preventing high level lymphokine synthesis by activated T cells, will be useful in situations of tissue, skin and organ transplantation and in graft-versus-host disease (GVHD). For example, blockage of T cell function should result in reduced tissue destruction in tissue transplantation. Typically, in tissue transplants, rejection of the transplant is initiated through its recognition as foreign by T cells, followed by an immune reaction that destroys the transplant. The administration of a molecule which inhibits or blocks interaction of a B7 lymphocyte antigen with its natural ligand(s) on immune cells (such as a soluble, monomeric form of a peptide having B7-2 activity alone or in conjunction with a monomeric form of a peptide having an activity of another B lymphocyte antigen (*e.g.*, B7-1, B7-3) or blocking antibody), prior to transplantation can lead to the binding of the molecule to the natural ligand(s) on the immune cells without transmitting the corresponding costimulatory signal. Blocking B lymphocyte antigen function in this matter prevents cytokine synthesis by immune cells, such as T cells, and thus acts as an immunosuppressant. Moreover, the lack of costimulation may also be sufficient to energize the T cells, thereby inducing tolerance in a subject. Induction of long-term tolerance by B lymphocyte antigen-blocking reagents may avoid the necessity of repeated administration of these blocking reagents. To achieve sufficient immunosuppression or tolerance in a subject, it may also be necessary to block the function of B lymphocyte antigens.

The efficacy of particular blocking reagents in preventing organ transplant rejection or GVHD can be assessed using animal models that are predictive of efficacy in humans.

Examples of appropriate systems which can be used include allogeneic cardiac grafts in rats and xenogeneic pancreatic islet cell grafts in mice, both of which have been used to examine the immunosuppressive effects of CTLA4Ig fusion proteins *in vivo* as described in Lenschow *et al.*, Science 257:789-792 (1992) and Turka *et al.*, Proc Natl Acad Sci USA, 89:11102-11105 (1992). In addition, murine models of GVHD (see Paul ed., FUNDAMENTAL IMMUNOLOGY, Raven Press, New York, 1989, pp. 846-847) can be used to determine the effect of blocking B lymphocyte antigen function *in vivo* on the development of that disease.

Blocking antigen function may also be therapeutically useful for treating autoimmune diseases. Many autoimmune disorders are the result of inappropriate activation of T cells that are reactive against self tissue and which promote the production of cytokines and auto-antibodies involved in the pathology of the diseases. Preventing the activation of autoreactive T cells may reduce or eliminate disease symptoms. Administration of reagents which block costimulation of T cells by disrupting receptor:ligand interactions of B lymphocyte antigens can be used to inhibit T cell activation and prevent production of auto-antibodies or T cell-derived cytokines which may be involved in the disease process. Additionally, blocking reagents may induce antigen-specific tolerance of autoreactive T cells which could lead to long-term relief from the disease. The efficacy of blocking reagents in preventing or alleviating autoimmune disorders can be determined using a number of well-characterized animal models of human autoimmune diseases. Examples include murine experimental autoimmune encephalitis, systemic lupus erythematosus in MRL/lpr/lpr mice or NZB hybrid mice, murine autoimmune collagen arthritis, diabetes mellitus in NOD mice and BB rats, and murine experimental myasthenia gravis (see Paul ed., FUNDAMENTAL IMMUNOLOGY, Raven Press, New York, 1989, pp. 840-856).

Upregulation of an antigen function (preferably a B lymphocyte antigen function), as a means of up regulating immune responses, may also be useful in therapy. Upregulation of immune responses may be in the form of enhancing an existing immune response or eliciting an initial immune response. For example, enhancing an immune response through stimulating B lymphocyte antigen function may be useful in cases of viral infection. In addition, systemic vital diseases such as influenza, the common cold, and encephalitis might be alleviated by the administration of stimulatory forms of B lymphocyte antigens systemically.

Alternatively, anti-viral immune responses may be enhanced in an infected patient by removing T cells from the patient, costimulating the T cells *in vitro* with viral antigen-pulsed APCs either expressing a peptide of the present invention or together with a stimulatory form of a soluble peptide of the present invention and reintroducing the *in vitro* activated T cells
5 into the patient. Another method of enhancing anti-viral immune responses would be to isolate infected cells from a patient, transfect them with a nucleic acid encoding a protein of the present invention as described herein such that the cells express all or a portion of the protein on their surface, and reintroduce the transfected cells into the patient. The infected cells would now be capable of delivering a costimulatory signal to, and thereby activate, T
10 cells *in vivo*.

In another application, up regulation or enhancement of antigen function (preferably B lymphocyte antigen function) may be useful in the induction of tumor immunity. Tumor cells (*e.g.*, sarcoma, melanoma, lymphoma, leukemia, neuroblastoma, carcinoma) transfected with a nucleic acid encoding at least one peptide of the present invention can be administered to a
15 subject to overcome tumor-specific tolerance in the subject. If desired, the tumor cell can be transfected to express a combination of peptides. For example, tumor cells obtained from a patient can be transfected *ex vivo* with an expression vector directing the expression of a peptide having B7-2-like activity alone, or in conjunction with a peptide having B7-1-like activity and/or B7-3-like activity. The transfected tumor cells are returned to the patient to
20 result in expression of the peptides on the surface of the transfected cell. Alternatively, gene therapy techniques can be used to target a tumor cell for transfection *in vivo*.

The presence of the peptide of the present invention having the activity of a B lymphocyte antigen(s) on the surface of the tumor cell provides the necessary costimulation signal to T cells to induce a T cell mediated immune response against the transfected tumor
25 cells. In addition, tumor cells which lack MHC class I or MHC class II molecules, or which fail to reexpress sufficient amounts of MHC class I or MHC class II molecules, can be transfected with nucleic acid encoding all or a portion of (*e.g.*, a cytoplasmic-domain truncated portion) of an MHC class I α chain protein and β_2 microglobulin protein or an MHC class II a chain protein and an MHC class II β chain protein to thereby express MHC class I or MHC
30 class II proteins on the cell surface. Expression of the appropriate class I or class II MHC in

conjunction with a peptide having the activity of a B lymphocyte antigen (e.g., B7-1, B7-2, B7-3) induces a T cell mediated immune response against the transfected tumor cell.

Optionally, a gene encoding an antisense construct which blocks expression of an MHC class II associated protein, such as the invariant chain, can also be cotransfected with a DNA

5 encoding a peptide having the activity of a B lymphocyte antigen to promote presentation of tumor associated antigens and induce tumor specific immunity. Thus, the induction of a T cell mediated immune response in a human subject may be sufficient to overcome tumor-specific tolerance in the subject.

The activity of a polypeptide of the invention may, among other means, be measured
10 by the following methods: Suitable assays for thymocyte or splenocyte cytotoxicity include, without limitation, those described In: CURRENT PROTOCOLS IN IMMUNOLOGY. Coligan *et al.*, eds. Greene Publishing Associates and Wiley-Interscience (Chapter 3, Chapter 7); Herrmann *et al.*, *Proc Natl Acad Sci USA* 78:2488-2492, 1981; Herrmann *et al.*, *J Immunol* 128:1968-1974, 1982; Handa *et al.*, *J Immunol* 135:1564-1572, 1985; Takai *et al.*, *J Immunol*
15 137:3494-3500, 1986; Takai *et al.*, *J Immunol* 140:508-512, 1988; Herrmann *et al.*, *Proc Natl Acad Sci USA* 78:2488-2492, 1981; Herrmann *et al.*, *J Immunol* 128:1968-1974, 1982; Handa *et al.*, *J Immunol* 135:1564-1572, 1985; Takai *et al.*, *J Immunol* 137:3494-3500, 1986; Bowman *et al.*, *J Virology* 61:1992-1998; Takai *et al.*, *J Immunol* 140:508-512, 1988; Bertagnolli *et al.*, *Cell Immunol* 133:327-341, 1991; Brown *et al.*, *J Immunol* 153:3079-3092,
20 1994.

Assays for T-cell-dependent immunoglobulin responses and isotype switching (which will identify, among others, proteins that modulate T-cell dependent antibody responses and that affect Th1/Th2 profiles) include, without limitation, those described in: Maliszewski, *J Immunol* 144:3028-3033, 1990; and Mond and Brunswick In: CURRENT PROTOCOLS IN
25 IMMUNOLOGY. Coligan *et al.*, (eds.) Vol 1 pp. 3.8.1-3.8.16, John Wiley and Sons, Toronto 1994.

Mixed lymphocyte reaction (MLR) assays (which will identify, among others, proteins that generate predominantly Th1 and CTL responses) include, without limitation, those described In: CURRENT PROTOCOLS IN IMMUNOLOGY. Coligan *et al.*, eds. Greene Publishing
30 Associates and Wiley-Interscience (Chapter 3, Chapter 7); Takai *et al.*, *J Immunol*

137:3494-3500, 1986; Takai *et al.*, *J Immunol* 140:508-512, 1988; Bertagnolli *et al.*, *J Immunol* 149:3778-3783, 1992.

Dendritic cell-dependent assays (which will identify, among others, proteins expressed by dendritic cells that activate naive T-cells) include, without limitation, those described in:

- 5 Guery *et al.*, *J Immunol* 134:536-544, 1995; Inaba *et al.*, *J Exp Med* 173:549-559, 1991; Macatonia *et al.*, *J Immunol* 154:5071-5079, 1995; Porgador *et al.*, *J Exp Med* 182:255-260, 1995; Nair *et al.*, *J Virol* 67:4062-4069, 1993; Huang *et al.*, *Science* 264:961-965, 1994; Macatonia *et al.*, *J Exp Med* 169:1255-1264, 1989; Bhardwaj *et al.*, *J Clin Investig* 94:797-807, 1994; and Inaba *et al.*, *J Exp Med* 172:631-640, 1990.

- 10 Assays for lymphocyte survival/apoptosis (which will identify, among others, proteins that prevent apoptosis after superantigen induction and proteins that regulate lymphocyte homeostasis) include, without limitation, those described in: Darzynkiewicz *et al.*, *Cytometry* 13:795-808, 1992; Gorczyca *et al.*, *Leukemia* 7:659-670, 1993; Gorczyca *et al.*, *Cancer Res* 53:1945-1951, 1993; Itoh *et al.*, *Cell* 66:233-243, 1991; Zacharchuk, *J Immunol* 145:4037-4045, 1990; Zamai *et al.*, *Cytometry* 14:891-897, 1993; Gorczyca *et al.*, *Internat J Oncol* 1:639-648, 1992.

- Assays for proteins that influence early steps of T-cell commitment and development include, without limitation, those described in: Antica *et al.*, *Blood* 84:111-117, 1994; Fine *et al.*, *Cell Immunol* 155: 111-122, 1994; Galy *et al.*, *Blood* 85:2770-2778, 1995; Toki *et al.*,
20 *Proc Nat Acad Sci USA* 88:7548-7551, 1991.

Other Activities

- A polypeptide of the invention may also exhibit one or more of the following additional activities or effects: inhibiting the growth, infection or function of, or killing, infectious agents, including, without limitation, bacteria, viruses, fungi and other parasites;
25 effecting (suppressing or enhancing) bodily characteristics, including, without limitation, height, weight, hair color, eye color, skin, fat to lean ratio or other tissue pigmentation, or organ or body part size or shape (such as, for example, breast augmentation or diminution, change in bone form or shape); effecting biorhythms or circadian cycles or rhythms; effecting the fertility of male or female subjects; effecting the metabolism, catabolism, anabolism,
30 processing, utilization, storage or elimination of dietary fat, lipid, protein, carbohydrate,

vitamins, minerals, cofactors or other nutritional factors or component(s); effecting behavioral characteristics, including, without limitation, appetite, libido, stress, cognition (including cognitive disorders), depression (including depressive disorders) and violent behaviors; providing analgesic effects or other pain reducing effects; promoting differentiation and
5 growth of embryonic stem cells in lineages other than hematopoietic lineages; hormonal or endocrine activity; in the case of enzymes, correcting deficiencies of the enzyme and treating deficiency-related diseases; treatment of hyperproliferative disorders (such as, for example, psoriasis); immunoglobulin-like activity (such as, for example, the ability to bind antigens or complement); and the ability to act as an antigen in a vaccine composition to raise an immune
10 response against such protein or another material or entity which is cross-reactive with such protein.

The present invention is not to be limited in scope by the specific embodiments described herein. Indeed, various modifications of the invention in addition to those described herein will become apparent to those skilled in the art from the foregoing description and
15 accompanying figures. Such modifications are intended to fall within the scope of the appended claims.

The contents of all references, patents and published patent applications cited throughout this application are hereby incorporated by reference.

EQUIVALENTS

20 It is to be understood that while the invention has been described in conjunction with the detailed description thereof, the foregoing description is intended to illustrate and not limit the scope of the invention, which is defined by the scope of the appended claims. Other aspects, advantages, and modifications are within the scope of the following claims.